

A review of organic waste enrichment for inducing palatability of black soldier fly larvae: Wastes to valuable resources

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Abstract

The increase of annual organic wastes generated worldwide has become a major problem for many countries since the mismanagement could bring about negative effects on the environment besides, being costly for an innocuous disposal. Recently, insect larvae have been investigated to valorize organic wastes. This entomoremediation approach is rising from the ability of the insect larvae to convert organic wastes into its biomass via assimilation process as catapulted by the natural demand to complete its lifecycle. Among the insect species, black soldier fly or *Hermetia illucens* is widely researched since the larvae can grow in various environments while being saprophagous in nature. Even though black soldier fly larvae (BSFL) can ingest various decay materials, some organic wastes such as sewage sludge or lignocellulosic wastes such as waste coconut endosperm are destitute of decent nutrients that could retard the BSFL growth. Hence, blending with nutrient-rich low-cost substrates such as palm kernel expeller, soybean curd residue, etc. is employed to fortify the nutritional contents of larval feeding substrates prior to administering to the BSFL. Alternatively, microbial fermentation can be adopted to breakdown the lignocellulosic wastes, exuding essential nutrients for growing BSFL. Upon reaching maturity, the BSFL can be harvested to serve as the protein and lipid feedstock. The larval protein can be made into insect meal for farmed animals, whilst the lipid source could be extracted and transesterified into larval biodiesel to cushion the global energy demands. Henceforth, this review presents the influence of various organic wastes introduced to feed BSFL, targeting to reduce wastes and producing biochemicals from mature larvae through entomoremediation. Modification of recalcitrant organic wastes via fermentation processes is also unveiled to ameliorate the BSFL growth. Lastly, the sustainable applications of harvested BSFL biomass are as well covered together with the immediate shortcomings that entail further researches.

Keywords: Black soldier fly larva; Waste management; Blended substrate; Fermentation; Biochemical

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1. Introduction

Lately, the quantity of organic wastes generated worldwide continues to increase substantially in order to enliven human consumptions. The discarded organic wastes include food wastes and undesired byproducts from various industries, namely, sewage sludge from wastewater treatment plants, animal manure from agricultural farms, soybean curd residue from tofu manufacturing process, etc. Globally, approximate 2.01 billion metric tons/year of municipal solid wastes is generated, and this amount is expected to reach 3.40 billion metric tons/year by 2050 (Ellis, 2018). Worse still, the generated organic wastes have inevitably led to large carbon dioxide emissions, i.e., 1.6 billion metric tons of carbon dioxide equivalents worldwide. The emission is unabatedly projected to increase to 2.6 billion metric tons by year 2050 (Ellis, 2018). Furthermore, the mismanagement of organic wastes can as well contribute to the broad environmental menaces and economic woes (Ferronato and Torretta, 2019). Thus, a proper planning of waste disposal and reduction is required in a bid to manage the enormous organic wastes safely and sustainably with a minimum spewing of greenhouse gasses. In this regard, the employment of insect larvae for organic wastes reduction has been proven effective and environmentally friendly since the larvae can ingest organic wastes, transforming it into larval biomass through the assimilation process without harming humans and the surrounding environments. Black soldier fly or *Hermetia illucens* is considered as an ideal insect species since the larvae (BSFL) can bioconvert various decay matters, survive among variety of surrounding conditions, inhibit the untoward microbe's growths, and most importantly, the adult is not a pest (Caruso et al., 2014; Nguyen et al., 2013; Tomberlin and Cammack, 2017; Yu et al., 2011). The essential component in larval feeding substrates is protein, in which could have a significant positive impact on the BSFL development to complete its lifecycle (Gold et al., 2020). Nevertheless, some organic wastes exploited as the feeding substrates, such as sewage sludge or lignocellulosic waste coconut endosperm, contain insufficient nutrients to support the BSFL growth (Leong et al., 2016; Mohd-Noor et al., 2017). The lignocellulosic wastes composing of lignin, cellulose and hemicellulose can significantly hinder the digestion process of BSFL, and subsequently retarding its growth measured in terms of slow larval development time or small prepupal weight and size. Therefore, blending with nutrient-rich low-cost substrates (soybean curd residue, palm kernel expeller, etc.) or fermenting in the presence of exo-microbes (*Saccharomyces cerevisiae*,

Bacillus subtilis, *Lactobacillus buchneri*, etc.) via *in-situ* and *ex-situ* conditions can consequentially enrich the feeding substrate, prompting the palatability of BSFL. Once the BSFL mature, its biomass can be harvested to serve as the feedstock for protein, lipid and other biochemicals productions. The protein from BSFL biomass can be used to substitute the traditional aquatic meal and poultry feed, tapering off the cost of protein sources derived from soybean or fish meals. The lipid extracted from BSFL biomass is valuable for producing high quality biofuel to sate the rising demand of energy consumptions. Moreover, the utilization BSFL-based lipid as a sustainable raw material is virtually regarded as a new generation of biofuel production.

2. Current management of solid organic wastes

Today, various consumer products have been manufactured qualitatively and quantitatively in order to satiate the demands of increasing population in many ways. Along the manufacturing processes, vast organic waste materials have been generated in the form of by-products. This has directly given rise to the inevitable challenges for disposal as the organic fraction from solid waste stream can spew excessive greenhouse gases during degradation (Hoornweg and Bhada-Tata, 2012). The municipal solid wastes have reached 2.01 billion tons/year worldwide recently and it is projected to ratchet up to 2.59 billion tons/year by 2030 (Fig. 1a) (Kaza et al., 2018). The largest composition of municipal solid wastes is the organic waste materials, i.e., encompassing more than 44% (Fig. 1b) (Hoornweg and Bhada-Tata, 2012).

2.1 Major organic wastes

A major source of organic wastes is in the form of food waste. Globally, one-third of foods generated are wasted, i.e., about 1.3 billion tons/year of foods are not consumed as reported by FAO, 2020 (Fig. 2). In Malaysia, on average, a family will discard around 0.5-0.8 kg of foods everyday (Bong et al., 2017). Moreover, Malaysia was reported to have achieved about 15 000 tons/day of food wastes by the Statistics from Solid Waste Corporation of Malaysia (Hoornweg and Bhada-Tata, 2012). The enormous solid organic wastes can afflict human health, in addition to the basic services such as waste management, district facilities, water supply and transport basic structure in any country. Also, this is very costly to the municipal budget in managing and disposing the solid organic wastes (Liyala, 2011). Alas, as a consequence of solid organic wastes disposal, the emission of carbon dioxide gas was estimated had reached 1.6 billion tons in 2016 and this figure is expected to grow to 2.38

119 billion tons/year by 2050 if the disposal method is still through landfilling or open dumping
120 without a proper gas collection system (Kaza et al., 2018). The greenhouse gas emissions
121 from three different classes for solid waste disposal that have been adopted worldwide are
122 landfilling, incineration without energy recovery and other waste treatment methods. Among
123 the classes, 95% of greenhouse gas was emitted from landfilling disposal mode. The other
124 two classes had merely recorded a total of 5% of greenhouse gas emissions (Fig. 3) (EEA
125 Greenhouse gas data viewer., 2014) . This indicates that landfilling has incontrovertibly
126 crowded out as the major factor contributing to the global warming. To make matters worse,
127 the landfilled organics can as well release a large amount of methane gas, leading to the
128 infrared radiation absorption whilst accelerating the global warming and climate change
129 phenomena (Move for Hunger, 2015; Recycle Bank, 2006).

130 Apart from food wastes, sewage sludge, a by-product generated from biological
131 wastewater treatment plants, is also considered as a major solid organic waste material. The
132 productions of sewage sludge in Europe, United States and China had been reported to be 10
133 million tons, 49 trillion liters and 20 million tons/year, respectively. Locally in Malaysia,
134 sewage sludge is generated by around 3 million metric tons annually and expected to reach 7
135 million metric tons in the year of 2020 (Oladejo et al., 2019; Roslan et al., 2013). Although
136 sewage sludge contains some valuable nutrients, its utilizations such as reuse or disposal to
137 landfills, composting and storage are limited by the heavy metals and toxic components laden
138 into sewage sludge during the discharge from industries or traffic related pollutions (Mateo-
139 Sagasta et al., 2015). As of now, virtually, all countries are depending on landfilling to
140 manage the generated sewage sludge, e.g., EU-27, United States and China were disposing
141 12%, 30% and 80% of sewage sludge to the landfill sites, respectively. This traditional solid
142 waste management method is, indeed, not sustainable while enlarging the carbon footprint
143 insidiously (Mateo-Sagasta et al., 2015). In Malaysia, the estimated cost of management is
144 about US\$ 0.33 billion per year to dispose the sewage sludge due to its high contents of
145 pathogens, micro-pollutants, heavy metals and other hazardous substances, depending on the
146 origin of wastewaters (Kadir and Velayutham, 1999).

147 In addition, other solid organic waste, namely, animal manure from agricultural farm,
148 has increased in terms of capacity to the tune of over 1500 million tons/year in the EU-27
149 alone (FAO, 2003). Consequentially, 10% of greenhouse gases, 65% of N₂O and 64% of NH₃
150 are emitted globally with the origin from agricultural activities, primarily via animal manure
151 productions (Gómez-Brandón et al., 2013; Steinfeld et al., 2006). The current management
152 strategy for animal manure is through land application, i.e., apportioned into the soil.

However, the major challenge is the presence of high level of nitrogen and phosphorus compounds contaminating the soil and later bringing about to the environmental pollution. Thus, the management of animal manure as well as agricultural waste via recent technology such as compaction or composting has considered. However, these treatment methods require a high expense (Szogi et al., 2015). Moreover, the mismanagement of animal manures can result in the emergence and spread of contagious diseases since the manures are generally hosting various dangerous and infectable microorganisms. The methane gas is also released from the fresh animal manure that has passed through the enteric fermentation of farmed animals; because of that, it intensifies the greenhouse gas emission. Meanwhile, the release of ammonia gas has mightily polluted the environment by causing Eutrophication to the natural water bodies (Aguirre-Villegas and Larson, 2017; Gómez-Brandón et al., 2013; Lim et al., 2016; Loyon, 2018; Malomo et al., 2018). The land application to dispose the generated animal manures will introduce excessive phosphorus sources into the soils, saturating the capacity of soils to retain phosphorus and subsequently, leaching the soluble phosphorus species via continuous surface runoff and erosion (Zhang and Schroder, 2014). Following the ammonia, the presence of phosphorus sources in natural water bodies will as well hasten the Eutrophication menace. Thus, to sum up, the solid organic wastes entail an inclusive management for the sake of mitigating the environmental risks and reducing the long-term costs of disposal (Attigbe et al., 2019).

2.2 Management of organic wastes

The traditional methods up to the recent approaches that have been employed to handle the solid organic wastes are landfilling, incineration or combustion, recovery and recycling, plasma gasification, composting, energy recovery and avoidance or waste minimization (Fig. 4). The broadly applied method to dispose the solid organic wastes and other garbage is dumping into landfill sites, i.e., encompassing over 60% of global waste disposal and 80% of was reduction in Malaysia, due to convenient and low-cost (Hoornweg and Bhada-Tata, 2012; Kaza et al., 2018). The procedure of landfilling method begins with solid wastes being buried beneath the trash layer. The organic fraction is then biodegraded under the aerobic condition until all the oxygen has depleted in the subsurface. Then, it will pave the way for the anaerobic biodegradation of remnant organics to transpire while spewing methane gas into atmosphere which is 25 times more powerful than CO₂ to cause global warming (Amritha and Anilkumar, 2016). In fact, landfilling has crowded out as the third largest source of methane emissions globally (Amritha and Anilkumar, 2016; Zhao et

al., 2019). Other problem relates to landfilling is a leachate production, carrying the soluble compounds from the site and contaminating the adjacent water sources, especially the groundwater. In accounting this unsustainable approach, Malaysia has pledged to decrease the disposal of solid wastes via landfilling to 65% by 2020 (Hoornweg and Bhada-Tata, 2012; Malek and Shaaban, 2008; Sauve and Van Acker, 2020). Other resemblance method is open dumping. The receivers of open dumping can be lands or water bodies in which occupying a fraction of 33% (Kaza et al., 2018). Similar to the landfilling, the open dumping can also accelerate the global warming due to the emission of greenhouse gases especially methane that is generated from decomposing of biodegradable organics under anaerobic condition (Couth and Trois, 2009; Isibika et al., 2019).

Besides landfilling, anaerobic digestion and composting are considered as the alternative ways to stabilize solid organic wastes. Anaerobic digestion is a process to decompose organic waste materials in the absence of oxygen, whilst composting is a process to promote decomposition of organic waste materials under the aerobic condition (Kaza et al., 2018). Indeed, composting has accounted 55% of solid organic wastes treatment worldwide. The product derived from both the anaerobic digestion and composting processes can be utilized as a fertilizer for agriculture or landscaping purposes (Aggelides and Londra, 2000; Cheng et al., 2007; Lim et al., 2016). Nevertheless, the treatment of solid organic wastes by both methods needs to be managed adequately since it can contribute to the environmental problems like landfilling (Pace et al., 2018). For instance, the application of digestate, a product after anaerobic digestion process, directly in land can give rise to the uncontrolled greenhouse gas emissions since degradable substrates, phytotoxins and methanogenic microbiota remain in the digestate. Thereby, continuing the anaerobic process to further decompose the remnant organics or volatile fatty acids into methane and other greenhouse gasses (Kirchmann and Bernal, 1997; Kirchmann and Lundvall, 1993).

Incineration is one of the rapid solid waste reduction techniques using oxygen for combusting during the process. The amount of solid wastes stabilization by incineration in worldwide is about 11.1%. In the case of China, the employment of incinerations to stabilize solid wastes had increased dramatically from 3.7 million tons to 61.7 million tons between 2003 and 2015, i.e., the later had amounted 32.5% of solid wastes reduction in China (Hong et al., 2017; National Bureau of Statistics of the People's Republic of China, 2015). Even though incineration can generate heat and electrical energy from solid wastes while reducing a large volume of wastes, it requires professional managements in dealing with air pollutants emission such as CO₂, NO_x and ash residue during as well as after the incineration process,

respectively (Beylot et al., 2018; Wang et al., 2018). Moreover, incineration also incurs investment to compensate the potential losses from incineration, e.g., decreasing of biodiversity, injuring public health and accessing of land (Wang et al., 2018).

The sewage sludge disposal through sewer (underground closed pipes) by water carriage system, i.e., water will carry sewage sludge to the disposal place, has been exploited globally at the present since this new method is suitable for the management of sewage sludge. In addition, the maintenance cost is not expensive while utilizing a small footprint of lands since most of the pipes are hidden underground without directly interfering the land developments. Nonetheless, this system requires a high initial cost for investment and 99% of water carrier will be eventually wasted (Engineering Articles, 2015).

In considering the sustainability aspects of currently employed techniques to reduce the solid organic wastes, it seems precarious and not promising for long-term applications. Thus, biological conversion or bioconversion of solid organic wastes by insect larvae has been investigated recently to overcome the setbacks experienced by the currently employed techniques. The outcomes concluded from the studies had confirmed the feasibility of a novel bioconversion technique to stabilize the waste organics via valorization, while benefitting the environment. Various insect or its larval species had been selected to bioconvert organic wastes into valuable biomass such as yellow meal worm, i.e., *Tenebrio molitor* L. in Coleoptera order, black soldier fly, i.e., *Hermetia illucens* L. in Diptera order, face fly (*Musca autumnalis* L.) , flesh fly (*Sarcophaga carnaria* L.) and house fly (*Musca domestica* L.) (Čičková et al., 2015; Wang et al., 2017b). The yellow meal worm larvae contain about 23% - 47% of fat content and have the ability to consume decayed vegetable as a feeding substrate (Alves et al., 2016; Veldkamp et al., 2012; Zheng et al., 2013). Apart from that, house fly larvae also can grow in solid organic waste materials and manure diets (Čičková et al., 2015). Even though house fly presents a rapid reproduction rate and easy for rearing, it is a pest and can widely spread diseases and parasites (Förster et al., 2007; Förster et al., 2009; Hogsette and Farkas, 2000). Next, face fly can grow by feeding organic substrates, especially cattle manure. However, face fly is a threat for cattle and horse since it can transmit diseases to these animals such as pink eye or thelaziasis. The flesh fly can be also reared with waste organic feeds; but it is difficult to identify its larval and adult stages, leading to the obstruction amidst experiments (Čičková et al., 2015). Among all the insect species, black soldier fly larvae (BSFL) is considered as a potential insect since it can consume a variety of solid organic waste materials. Also, the mature BSFL contain about 20% - 40% of fat content. Thus, the BSFL biomass can be exploited as the protein and lipid feedstock for

poultry feed and biodiesel production, respectively (Dierenfeld and King, 2008; Oonincx et al., 2015). The significant strengths of BSFL are not only it can assist to control the oviposition of house flies that can inflict human and animals health, but also the adults are not a pest and the larvae are saprophagous in nature, capable of valorizing various and large amount of solid organic wastes (Čičková et al., 2015).

3. Lifecycle of black soldier fly

Black soldier fly (BSF) or *Hermetia illucens* is a common tropical and sub-tropical insect. This species has gained increasing interests among the fervent researchers recently to serve as the potential feedstock for larval lipid and protein productions. During the growing stage, the larvae of black soldier fly (BSFL) can accumulate various essential biochemical contents within the structural space between its organs for the uses amidst non-feeding period in its lifecycle, i.e., to undergo pupation (Manzano-Agugliaro et al., 2012). Moreover, the BSFL biomass also has higher content of saturated fatty acids as compared with other species of insects (Ramos-Bueno et al., 2016). Apart from being saprophagous and polyphagous in assimilating myriad organic waste materials such as fruit and vegetable waste or animal manure (Nguyen et al., 2013), the BSFL also can live in various environments, inhibit the growth of untoward microbes while serving as an animal feed and the adult fly is not a pest in nature (Caruso et al., 2014; Yu et al., 2011). The lifecycle of BSF takes about 40 to 50 days (Fig. 5). It begins with the female fly ovipositing eggs near the decomposing organic matters, rendering as a food source for the neonates of BSF. Then, the female fly will die thereafter its energy is exhausted. After about 4 days, the eggs will hatch and the neonates of BSFL will emerge. The larvae which have a creamy color will ingest the surrounding decomposed organics as its food source. This larval stage is the only feeding period for BSF that will extend until reaching a 5th instar stage, i.e., approximately 4 weeks, depending on the quality and availability of organics that can be ingested. Thereafter, the BSFL will undergo eclosion into the prepupae in achieving its 6th instar stage, viz. the last stage of larval form when its light brown color is darkened. During this period as well, the BSFL will stop ingesting organics and its mouthpart will be transformed into hook-shaped structure in aiding the prepupae for moving away from organics to ensconce in a dry place for pupation (Dortmans et al., 2017). The pupa will finally transform into a fly whereby the mature BSF will start spreading its wings and flying off from the cocoon. The pupation is the last eclosion process for BSF and it will consume ca. a week. The emerged BSF will live averagely for 4-6 days for mating and ovipositing eggs in continuing its lifecycle.

4. Feeding substrates for BSFL

The prime characteristic of BSF is the larvae can ingest-cum-valorize various organic materials inclusive of decomposable byproducts and wastes for growth until reaching prepupae. In fact, the larval stage is the only feeding duration in BSF lifecycle, i.e., the BSFL need to accumulate sufficient nutrients such as lipids and proteins prior to the pupation and subsequent emergence into adult flies. Thus, the quality of larval feeding substrates especially protein and carbohydrate contents can significantly affect the BSFL growths, organic bioconversion efficiencies, prepupal weights and nutritional contents of mature BSFL in which are generally consisting of approximately 40% of larval protein and 30% of larval lipid (dry weight basis) (Barragan-Fonseca et al., 2017; Kinasih et al., 2018). Moreover, the presence of large amount of larval feeding substrates could assist the BSFL to partially overcome the low quality of nutrients composition. Because of that, shortening the larval development time even small pupal sizes were eventually harvested (Kinasih et al., 2018). The feeding substrates for BSFL can be conventionally categorized into 2 types. The simplest larval feeding substrate consists of a single organic material and it is used directly for feeding of BSFL upon receiving. Conditioning of two or more single substrates via blending or fermenting through inoculation by the various microorganisms has as well been exploited, targeting to offset the shortcomings suffered from the use of some single substrates to grow BSFL.

4.1 Single substrate

There were many completed studies demonstrating the administration of various single substrates to feed BSFL. The commonly studied single substrates were chicken feed, animal manure, food waste, fruit and vegetable residue and sludge. In general, the growth of BSFL in concert with larval body nutritional contents vary with the type of substrates having been ingested during the rearing period. The BSFL body nutritional contents encompass proteins, minerals, amino acids, fats, etc. with the alimentation values directly depending on the physical properties and chemical compositions of the feeding substrates (Table 1) (Kinasih et al., 2018). This section reviews the impacts of feeding BSFL with various single substrates on larval development time from neonates to the first prepupa emergence, prepupal weight as well as harvested BSFL biomass nutritional composition (Table 2).

Among all the single substrates studied thus far, the use of chicken feed had given rise to the shortest larval development time as compared with animal manure, restaurant waste

and fruit and vegetable residue. The rearing period prior to the mature BSFL harvesting only entailed 12 days in which the first prepupa could be observed (Spranghers et al., 2017). The highest total larval biomass was also attained while using the chicken feed to grow BSFL. Kinasih et al. (2018) had also reported that the BSFL would require about 20 days for the first prepupa to emerge while controlling the larval feeding rate at 100 mg chicken feed/larva/day (typical feeding rate accepted by many researchers). The highest prepupal weight was recorded at approximately 130 mg/larva using this feeding control. It could be concluded that the chicken feed contained sufficient nutrients to enhance the palatability of BSFL in promoting its growth. Accordingly, it was found that the protein content in chicken feed was measured at about 175 g/kg dry weight (~18%) in which playing a pivotal role to spur the BSFL growth at the shortest time (Li et al., 2012; Spranghers et al., 2017; Tschirner and Simon, 2015). However, using the chicken feed for growing BSFL and later feeding the farmed chicken with mature BSFL is not an economical approach; unless, the harvested larval biomass has other commercial uses. In this regard, other low-cost single substrates were exploited to grow BSFL in order to curtail the production cost for producing BSFL biomass while valorizing the organics.

Besides chicken feed, animal manure is the next palatable substrate for BSFL feeding since it can as well spur the larval growth and body nutritional content comparably. Shumo et al. (2019) had revealed that the BSFL fed with chicken manure would obtain 41.1% of larval crude protein which was in conformity with the reported works by Sheppard et al. (1994), accentuating 42% of larval crude protein was measured while using a similar substrate. In addition, the harvested BSFL biomass was also found to contain high values of calcium mineral, i.e., 3.2 g/kg dry weight, and ash, i.e., 9.3 g/kg dry weight. The epidermis layer or outer layer of BSFL's skin could accumulate calcium mineral in the form of calcium carbonate, leading to the high level of calcium and ash contents in prepupae when early fed with chicken manure (Shumo et al., 2019). Moreover, some of the calcium and ash contents may be lost in the form of exuviae when the BSFL were undergoing stepwise eclosions in their early instars before harvesting. In other study, Newton et al. (2005) had analyzed the larval phosphorus content and found that the level was higher when the BSFL were fed with poultry manure as opposed to swine manure. Evidently, the different substrates, even in the form of manure, can significantly alter the BSFL body nutritional composition. For comparison, Kinasih et al. (2018) fed the BSFL with horse manure at the feeding rate of 100 mg/larva/day. The first prepupa was found emerging about 25 days later with the prepupal weight of merely 25 mg/larva, which was significantly lower than the BSFL having been fed

with chicken feed (130 mg/larvae). Also, the BSFL generally entailed longer rearing periods for emerging into prepupae when fed with animal manure than chicken feed. This was plausibly stemming from the destitute nutritional content of animal manure in which the BSFL would intrinsically ingest more substrate amount to accumulate minimum nutrients for eclosion into prepupae (Barry, 2004; Lee et al., 2004; Nijhout, 2003; Simpson et al., 2006; Wright et al., 2003); thereby, prolonging the rearing period. Furthermore, ur Rehman et al. (2017a) had found that, it was best if the BSFL feeding substrate was comprising of high total organic carbon. Although, the daily manure fulfills the larval diet need, the conversion efficiency into BSFL biomass is still low due to the presence of large amount of lignin, cellulose, hemicellulose biopolymers. These are not facilely digested biopolymers to BSFL and will ubiquitously lead to slow growth and small prepupae (Lalander et al., 2019).

The utilization of restaurant waste and fruit and vegetable residue as the BSFL feeding substrates had been investigated lately by many researchers. Spranghers et al. (2017) presented that the rearing period for BSFL fed with restaurant waste was 19 days for the first larva to emerge as the prepupa. The duration was shortened to 15 days when the larvae were administered with fruit and vegetable residue. The slow growth was primarily due to the presence of grease covering the restaurant waste, leading to the difficulty for BSFL to digest and convert the greasy waste into its body weight. Hence, the BSFL consumed more time for growing and developing into prepupae (Barry, 2004; Spranghers et al., 2017). Conversely, the fruit and vegetable residue was generally free from grease, oil and fat, favoring the physiological growth of BSFL. Apart from that, the harvested prepupae initially given with fruit and vegetable residue possessed a significantly higher ash content than the restaurant waste, namely, 96 and 27 g/kg dry weight, respectively. In the case of larval protein content, the BSFL could separately garner to the tune of 431 and 399 g/kg dry weight when fed with restaurant waste and fruit and vegetable residue, respectively (Spranghers et al., 2017). The higher larval ash content was possibly due to the presence of more minerals in fruit and vegetable residue than restaurant waste. Indeed, the presence of remnant meat composition in the restaurant waste could be the best justification of higher larval protein content than when fed with fruit and vegetable residue. Nevertheless, the difference in terms of larval protein contents was merely between approximately 43% and 40% while employing the restaurant waste and fruit and vegetable residue, respectively. Indeed, the use of fruit and vegetable residue to grow BSFL would enhance the larval mineral content (evidenced by high ash content), making the harvested larval biomass more suitable to serve as an animal feed. Later, Lalander et al. (2019) studied the application of fruit and vegetable residue as a feeding

substrate for BSFL and found that the prepupal weight of 218 mg/larva could be obtained when the rearing period was extended to 28 days. By using a conventional BSFL feeding rate of 100 mg/larva/day, Kinasih et al. (2018) revealed that the development time to reach prepupal stage was 25 days when fed with fruit and vegetable residue. Subsequently, by administering a fruit residue alone to BSFL, Leong et al. (2016) showed that a high growth rate could be attained since the fruit residue was consisting of high volatile solids, leading to the large larval size. However, the low protein in fruit residue had inevitably caused slow eclosion into prepupae. Concisely, the presence of high volatile solids and protein in larval diet, i.e., when employing a fruit and vegetable residue to grow BSFL, is the key parameter contributing to the high conversion efficiency into larval biomass and hastening the larval development into prepupae (Lalander et al., 2019). On the other hand, the utilization of restaurant waste to rear BSFL could be possibly improved if the excessive grease, fat and oil are skimmed prior to the BSFL feeding.

Next, the sludge from secondary wastewater treatment plants had also been exploited for BSFL development; whilst focusing on the sludge reduction via larval valorization. Nevertheless, the BSFL were found requiring a long development time of up to 39 days for the first prepupa to emerge. Worse still, the emerged sludge-fed prepupa was smaller in size (about 70 mg/larva) in comparison with using chicken feed, animal manure, restaurant waste or fruit and vegetable residue (Lalander et al., 2019). As a positive aspect, the employment of undigested sludge was found to reduce the larval development time to 30 days and double the larval size to 145 mg/larva. Leong et al. (2016) had associated the slow growth of BSFL while being fed with sewage sludge was due to the presence of inadequate volatile solids and protein contents in sludge. Consequentially, the small prepupal size would largely hinder the fertility of adult flies to reproduce, disrupting its lifecycle and later, its potential application to valorize sludge (Kinasih et al., 2018).

4.2 Blended substrate

The employment of low-cost single substrates has undoubtedly encountered several disadvantages for the rearing of BSFL. For instance, these single substrates of organic wastes are generally destitute of essential nutrients such as protein to enhance the larval growth. Moreover, during the processes to generate the organic wastes, the recalcitrant components for larval digestion such as lignin, cellulose, and hemicellulose are concentrated; thereby, hindering the BSFL development upon feeding, whilst later producing a low-value larval biomass. In fact, the palatable nutritional compositions that can ease the BSFL digestion are

proteins, non-fibre carbohydrates and modest amount of lipids (Barragan-Fonseca et al., 2018; Beniers and Graham, 2019; Casartelli et al., 2019; Lalander et al., 2019). The usual organic wastes administered for BSFL rearing are dairy manure, waste coconut endosperm and sludge since these wastes have been produced in humorous amounts as the by-products from industries and agricultural activities. However, the waste properties are improper for the BSFL development; and blending with other waste substrates is viewed as a potential option to fortify the diet for BSFL alimentation. Ideally, blending could improve the nutritional balance in terms of C/N ratios and buffer capacities (pH) that are essential for the enhancement of co-digestion efficiencies by BSFL (Anjum et al., 2016; Li et al., 2009). The conventional nutrient-rich substrate used for blending is soybean curd residue (SCR) in which it is a by-product derived from soy milk or tofu productions, the major surplus organic waste from soybean industries. Worse still, the disposal of SCR could increase the environmental impacts, particularly the release of greenhouse gasses (Li et al., 2013). Hence, using the SCR as a blending substrate for BSFL could enhance the efficiency of bioconversions due to a better nutritional balance in blended larval substrate (Table 2). Another potential substrate for blending is a palm kernel expeller which is the by-product generated from the palm oil extraction process. Although palm kernel expeller has a sufficient nutrient, i.e., containing a high level of crude protein (approximately 17%), the exploitations as a co-substrate for BSFL feeding have still not been documented as opposed to SCR that possesses a crude protein of more than 25% (Li et al., 2013).

The increasing demand for milk consumption has directly given rise to the excessive dairy manure generation from unplanned farming, which brings various environmental problems, namely, unpleasant odors, water pollutions, spreading of diseases, etc. (Aguirre-Villegas and Larson, 2017; Lim et al., 2016). To mitigate those issues, the BSFL have been employed to convert this organic waste. However, this larval valorization approach is very slow and not promising since the dairy manure consists of primarily lignin, cellulose and hemicellulose, which cannot be effectively digested by BSFL for growing, even the manure has a good buffer capacity (Li et al., 2016a; Mata-Alvarez et al., 2014; Wang et al., 2017a). Ur Rehman et al. (2017b) had shown that the 1000 BSFL fed with 1 kg of dairy manure required a long development time of 24 days for the first prepupal appearance. The blending of dairy manure with SCR for larval co-digestion was then compared since the SCR was generally rich in water insoluble nutrients such protein and fat that could significantly spur the BSFL growth. Besides, the low buffer capacity of SCR (pH~5.7) could also be offset by the better buffer capacity of dairy manure (pH~8.4) upon blending to suit the palatability of

BSFL (Li et al., 2016b; ur Rehman et al., 2017b). The results showed that the blended substrate between dairy manure and SCR could curtail the BSFL rearing period for emerging into prepupae and higher organic reduction rate could be measured as well with increasing of SCR proportions. The ratio of dairy manure to SCR at 1:4 had led to the shortest rearing time (21 days) for larval development into prepupae with the survival rate attained at 98.8% while reducing 75% of hemicellulose and 70% of cellulose. Other ratio with more dairy manure than SCR, for instance, 4:1, the BSFL could only reduce 45% of hemicellulose and 52% of cellulose after 22 days of rearing period (ur Rehman et al., 2017b), signifying the importance of substrates blending for an effective valorization of hemicellulose and cellulose. The plausible rationale was blending had balanced the nutrients requirement by BSFL for assimilating the blended substrate into its biomass at a more appropriate pH of feeding medium.

Apart from dairy manure, waste coconut endosperm has also been employed to feed BSFL since it is also an abundantly available organic waste derived from agriculture. The fresh coconut endosperm will lose its soluble components upon the coconut milk extraction, leaving behind a residue know as waste coconut endosperm. This organic waste mainly consists of lignocellulosic materials (30% of rough fibers) with low protein and fat contents, 5% and 9%, respectively. Thus, co-digestion of blended substrates is perceived as an alternative way to enhance the nutrients of waste coconut endosperm prior to feeding to the BSFL to perform bioconversion into valuable larval biomass. Again, SCR had been exploited to ameliorate the protein content of waste coconut endosperm via blending as reported by Lim et al. (2019). Initially, feeding the BSFL with a waste coconut endosperm alone had led to the lowest larval weight gained of only 32.5 mg/larva. The insufficient protein content in waste coconut endosperm would consequently accelerate the pupation process, resulting in small prepupae formation. At the optimum blended ratio of 3:2 between waste coconut endosperm and SCR, the feeding of this blended substrate had permitted the BSFL to attain the highest weight of 67.5 mg/larva, twice the weight of BSFL fed with only waste coconut endosperm. The BSFL could as well amass the highest larval body lipid and protein at 39.2 and 14.5 mg/larva, respectively, while feeding with the optimum blended ratio substrate. In comparison with BSFL fed with a single substrate of waste coconut endosperm, only 17.2 mg lipid/larva and 4.1 mg protein/larva could be measured from the harvested larval biomass. Nevertheless, a further increase in protein content in blended substrate by increasing the proportion of SCR over waste coconut endosperm had significantly decreased the harvested BSFL weight. As the SCR would degrade faster than waste coconut endosperm naturally, the

formation of ammonium in the blended substrate from protein degradation would acidify the larval medium, debilitating its buffer capacity. Also, the ammonia gas exuded from ammonium would retard the BSFL development since it affected the larval digestion system, leading to a small larval weight gained (Lim et al., 2019; Tschirner and Simon, 2015).

Sewage sludge is a well known organic waste produced from biological wastewater treatment plants worldwide. The traditional disposal of sewage sludge into the landfill will incontrovertibly extend the carbon footprint while having an expensive cost to handle. Thus, exploiting the sewage sludge for bioconversion into BSFL biomass is an alternative approach for waste reduction. Popa and Green (2012) confirmed that the BSFL had a potential for executing biotransformation when fed with a municipal raw sewage sludge. Although the sewage sludge is laden with various heavy metals such as lead, nickel, etc., none of the metals had influenced the BSFL lifecycle conspicuously as confirmed by Diener et al. (2015). The heavy metals from the sludge may be accumulated in BSFL body but it might not contaminate the lipid that was extracted from harvested larvae, for instance (Cai et al., 2018). Since the sewage sludge is generally lacking protein and digestible carbon sources, a facile blending with nutrient-rich organic matters could plausibly promote the co-digestion of sewage sludge by BSFL. Leong et al. (2016) presented that the BSFL weight had initially shown a small increment when fed with sewage sludge; however, its weight decreased after 4 days, resulting in an overall negative growth rate, i.e., -0.2 ± 0.01 mg/larva/day. Correspondingly, Cai et al. (2018) had also reported a similar observation in which the BSFL recorded a negative growth weight of -1.25 mg/larva due to the presence of insufficient nutrients in sewage sludge; leading to a short period of pupation process with small attainable larvae. In addition, the authors had further investigated the blending of chicken manure and wheat bran into sewage sludge to enhance the efficiency of BSFL bioconversion process. The results showed that the larval weight gained was proportional to the increase of either chicken manure or wheat bran portion in the blended substrates. The gains were recorded at 1.25, 10, and 20 mg/larva when fed with blended sewage sludge to chicken manure ratios of 75:25%, 50:50% and 25:75%, respectively. In the case of wheat bran, the study was investigated only for one ratio, namely, 84:16% of sewage sludge to wheat bran, with a weight gained determined at 12.5 mg/larva. However, when feeding the BSFL with blended substrates consisting of sewage sludge, chicken manure and wheat bran at various ratios, e.g., 63:21:16%, 42:42:16% and 21:63:16%, respectively, those were found to be better than any two blended substrates. The weight gained was recorded to be at least 22.5 mg/larva for any three blended substrates. The highest value gained could reach 28.75 mg/larva for an

527 optimum blended proportion of sewage sludge:chicken manure:wheat bran of 21:63:16%.
528 The BSFL rearing period was also reduced to only 12-13 days while using the optimum three
529 blended substrates as opposed to 30 days when fed with the sewage sludge alone. From the
530 principal component analysis and Pearson's correlation analysis, the contents of
531 carbohydrate, potassium and nitrogen presented in blended substrates were found to be the
532 primarily factors that simultaneously affecting the BSFL weight gained. Recently, Norgren et
533 al. (2019) had investigated the use of bio-sludge from wastewater treatment of pulp and paper
534 industry (PPBS) as a BSFL feeding substrate. The general composition of bio-sludge consists
535 of 1.5%–8.3% of crude protein, 0.3%–3.3% of fat and 17%–40% of lignin. The prepupal
536 weight gained upon feeding with this PPBS substrate was found to merely 0.4 mg/larva.
537 Nevertheless, the prepupal weight increased slightly when other substrates were blended into
538 PPBS, namely, 0.6 mg/larva when blended with water as a free surface, 2.0 mg/larva when
539 blended with composted leachate, 3.5 mg/larva when blended with leachate and water as a
540 free surface and finally, 4.8 mg/larva when blended leachate as a free surface. Although
541 adding some materials into PPBS could increase the prepupal weight, the results had shown
542 that the BSFL final weight was still very low because PPBS mainly comprised lignocellulosic
543 which was difficult to be digested by BSFL (Norgren et al., 2019). Cai et al. (2018) had
544 vindicated that using multiple blended substrates as the feeds for BSFL could result in better
545 larval growth, growth rate and bioconversion efficiency than any low-cost single substrate or
546 blending of two different substrates due to the more balance diets could be obtained from
547 more blended substrates. Gold et al. (2020) had inclusively studied the multiple blended
548 substrates for BSFL feeding. The formulations were calculated based on the composition of
549 single substrates and the achievement ratio of protein to non-fibre carbohydrate at around 1:1.
550 The results demonstrated that even at low larval feeding rate of 25%, feeding with multiple
551 substrates could still proffer decent outputs such as an average survival rate of 99% and
552 average larval weight of 43.5 mg/larva; in comparison with single substrate achieving an
553 average survival rate of 95% and average larval weight of 40.1 mg/larva. Gold et al. (2020)
554 had compared the performances of BSFL in valorizing the blended mill by-product, human
555 faeces, cow manure and vegetable waste at 23:16:11:50% (F1) against blended mill by-
556 product, canteen waste and vegetable waste at 33:33:33% (F2). Owing to the presence of
557 canteen waste, the BSFL could accumulate 22.3% of lipid with F2 as opposed to 19% with
558 F1 and 19.6% of protein with F2 as opposed to 13.8% with F1. However, the larval fiber
559 contents were slightly lower with F1 (38.5%) than F2 (39.8%). Indeed, employing F1 as a
560 BSFL feed had presented the best results, namely, 99.8% of survival rate, 64.2 mg/larva of

larval weight, 64.1% of waste reduction and 31.8% of biomass conversion rate in comparison with other blended formulations. It could be noted that even the multiple blended substrates comprising of lower lipid, protein and non-fibre carbohydrate contents than lesser type of blended substrates, the BSFL conversion efficiency and larval development were still better since the former were more nutritionally balance for BSFL.

4.3 Microbial fermented substrate

Another method to fortify the nutritional compositions of BSFL feeding substrates prior to the administrations is through microbial modification, i.e., by executing fermentation in waste biomasses or low-cost organics. The fermentation processes completed by various microorganisms can be specifically categorized into two types based on the inoculation modes. The *in-situ* fermentation transpires when the microorganisms are introduced to execute the fermentation process simultaneously with the valorization of organic substrates by BSFL. However, when the organic substrates are fermented early by the microorganisms before feeding to the BSFL is considered as *ex-situ* fermentation. In this case, the fermentation process is still ongoing during the larval feeding period. The presence of complex organic materials such as lignocelluloses from plant-based products in BSFL feeding substrates is generally difficult to be ingested since the larvae need to enter the epidermis layer of the plants prior to ingesting. Thus, microbial fermentation is deemed necessary to break down the complex components via hydrolysis while releasing myriad nutritional byproducts to spur the palatability of BSFL (Table 2) (Mohd-Noor et al., 2017; Wong et al., 2020).

4.3.1 In-situ fermentation

Among the microorganisms, *Saccharomyces cerevisiae*, a single cell yeast, had been employed to carry out *in-situ* fermentation in waste coconut endosperm before administering for BSFL feeding (Wong et al., 2020). The BSFL growth rate and waste-to-biomass conversion were found increasing with the increment of yeast concentrations, i.e., the highest values were achieved at 42.5 mg/larva/day and 11.5%, respectively, when the *in-situ* fermentation was carried out at the 2.5 wt% of yeast concentration. In comparison with the absence of yeast, the control waste coconut endosperm could only attain the larval growth rate of merely 3.25 mg/larva/day. Accordingly, the presence of yeast was propound could break down the carbohydrate compounds especially monosaccharides in waste coconut endosperm, leading to the better digestibility and assimilation of nutrients into BSFL bodies

(Wiedmeier et al., 1987; Yoon et al., 2003). However, the best lipid yield from harvest BSFL biomass, i.e., 49.4%, was attained when fed with 1.0 wt% of yeast concentration in waste coconut endosperm instead of 2.5 wt%. The transesterification of larval lipid permitted the biodiesel to contain a significant mixture of C12:0, C14:0, C16:0, and C18:1 (Table 3), indicating high in saturated fatty acids in which directly associated to the high in oxidative stability of produced biodiesel (Wong et al., 2020).

The *Bacillus subtilis*: S15, S16 and S19, isolated bacterial species from BSFL gut that could digest protein and organic phosphorus, had been exploited to inoculate chicken manure prior to feeding to the BSFL (Yu et al., 2010). The BSFL fed with any *B. subtilis in-situ* fermented chicken manure had resulted in higher prepupal weight and shorter development time in comparison with non-inoculated poultry manure. Nevertheless, among the *B. subtilis* species, the chicken manure inoculated with S15 had eventually engendered the highest prepupal weight of 94.6 mg/prepupa and shortest development time of 7.67 days. Moreover, Xiao et al. (2018) had also reported that the weight incremental rate of BSFL and material reduction rate by BSFL were increased by 15.9% and 40.5%, respectively, when fed with chicken manure initially inoculated with *B. subtilis* as opposed to control chicken manure. The symbiotic bacteria of *B. subtilis* could aid the assimilation process of BSFL to digest the non-digestible content, whilst providing the essential nutrients for BSFL growth as derived from fermentation process. Also, the presence of symbiotic *B. subtilis* would protect the growing BSFL from the surrounding risks such as parasitoids or pathogens (Douglas, 2015; Laughton et al., 2011). Other studies had also evidenced that the novel bacterial species isolated from immature black soldier fly could compost various organic materials, leading to the enhancement of BSFL development upon ingestion and more mature BSFL could be subsequently harvested (Ahmad et al., 2006; Bosch et al., 2014; Fitt and O'Brien, 1985; Xiao et al., 2018; Zheng et al., 2012b). Apart from that, besides treating sole chicken manure via *in-situ* fermentation, the *B. subtilis* had as well acted as an exogenous bacteria to colonize chicken manure blended with dairy manure prior to the BSFL feeding (ur Rehman et al., 2019). The dairy manure is usually rich in fibers such as the mixture of lignin, hemicellulose and cellulose that could hinder the BSFL assimilation for growth. Thus, the co-conversion with *B. subtilis* could assist the BSFL digestion process by modifying the fibers in terms of structure and chemicals (Li et al., 2016a; Masih-Das and Tao, 2018; ur Rehman et al., 2019). The mixing of dairy manure and chicken manure at the ratio of 2:3 had exhibited the best performances when simultaneously treated with *Bacillus* MRO₂ strain as a lignocellulosic degrading bacteria, leading to 99.07% of survival rate, 25.94 mg/larva of larval weight, 19

days of development time and 67.8% of lipid and 71.2% of protein utilizations for larval growth; as compared with the larval feeds inoculated with other bacteria strains or control feed that had attained the similar parameters at ca. 94.57%, 16.35 mg/larva, 20.58 days, 47.7% and 53.9%, respectively. In addition, the treated co-conversion substrate with *B. MRO₂* had also obtained high values of fiber reductions which were 72.96% for cellulose, 68.52% for hemicellulose and 32.86% for lignin. It was suggested that the exogenous bacteria could strengthen the gut microbiome of BSFL by modifying the ingested lignocelluloses to facilitate the BSFL digestion. Thereby, the reduction of animal manures went in tandem with the enhancement of BSFL development (ur Rehman et al., 2019).

The inoculation with *Lactobacillus buchneri* bacterial species in SCR had also been investigated by Somroo et al. (2019). The BSFL fed with *in-situ* fermented *L. buchneri* SCR had presented higher survival rate (98%), larval weight (34.7 mg/larva) and bioconversion rate (6.95%) and shorter development time (16.1 days) than the BSFL fed with fresh SCR, respectively recorded at 95.4%, 25 mg/larva, 5% and 17.7 days. It was rationalized that the co-digestion with *L. buchneri* played a significant role to support the BSFL adapting to the new surroundings and food sources. In this regard, the BSFL could benefit from the positive interactions in which more nutrients availability to enhance the BSFL growth, gut microbiota development and digestive enzyme production upon feeding with *in-situ* fermented SCR by *L. buchneri* (Engel and Moran, 2013; Kaltenpoth, 2009; Scott et al., 2008; Somroo et al., 2019; Teixeira et al., 2008; ur Rehman et al., 2017b; Yun et al., 2014). In accounting the nutritional contents of harvested BSFL biomass, the initial presence of *L. buchneri* had spurred the larval lipid and protein to 30% and 55.3%, respectively, as opposed to control SCR, without the prior *in-situ* fermentation, in which had recorded slightly lower contents, namely, 26.1% and 52.9%, respectively. Nevertheless, the addition of symbiotic *L. buchneri* had no effect on fatty acids composition in harvested BSFL and the main composition of fatty acids was consisting of saturated fatty acids which were C12:0, C16:0 and C14:0 (Table 3). Both the protein and fatty acids mixture derived from BSFL were later confirmed to be suitable serving as animal feed and biodiesel, respectively (Somroo et al., 2019).

Zheng et al. (2012a) had verified that the presence of mixed bacterial consortia, in the form of commercial product know as Rid-X, could aid the digestion of rice straw blended with restaurant waste in converting into BSFL biomass. The Rid-X was composed of natural bacteria that could degrade celluloses and hemicelluloses due to the presence of various enzymes exuded by bacteria such as cellulase, lipase, protease and amylase. The celluloses and hemicelluloses from *in-situ* fermented substrate could be reduced by 65% and 55%,

respectively, upon being valorized by BSFL as compared with only 27% and 32%, respectively, while using a control substrate. The lipid yield from 2000 larvae/batch was increased with the increase of Rid-X concentrations added into the blended rice straw (20%) and restaurant waste (80%), namely, approximately 32 g of lipid yield at 0.05 wt% of Rid-X and was increased to about 38 g of lipid yield at 0.4 wt% of Rid-X. Therefore, it could be concluded that the presence of more exo-bacteria could ultimately fasten the *in-situ* fermentation process and release more nutrients into BSFL feeding substrate to promote larval growth.

4.3.2 *Ex-situ* fermentation

The *ex-situ* approach via self-fermentation had been adopted by Mohd-Noor et al. (2017) to improve the nutritional characteristics of lignocellulosic biomass of waste coconut endosperm before administering to rear BSFL. The self-fermentation was associating to the ability of indigenous microorganisms to execute an intrinsic fermentation in organic materials over the time. The results had confirmed that the four weeks of fermentation's time were needed in order to release a maximum nutrient content from waste coconut endosperm, i.e., the highest total dissolved organic carbon concentration, especially organic acids, was measured at 70 ppm. Accordingly, the self-fermentation had mature once reaching four weeks in which the polysaccharides from waste coconut endosperm were significantly transformed into organic acids, softening the fiber property that was essential for maintaining the BSFL gut health (Caruso et al., 2014; Upadhaya et al., 2016). Consequentially, the BSFL achieved the highest growth of 35 mg/larva of weight gained and 2 mg/larva/day of growth rate. Furthermore, the BSFL also had accumulated the highest yields of lipid and protein at 57.95% and 15%, respectively, when fed with the week-4 self-fermented waste coconut endosperm. While using the fresh waste coconut endosperm (control) to feed the BSFL, the larval growths were only attained at approximately 22.5 mg/larva of weight gained and 1.5 mg/larva/day of growth rate with lipid and protein yields were found to be 20.70% and 12%, respectively, from harvested larval biomass (Mohd-Noor et al., 2017). Nevertheless, by increasing the self-fermentation period beyond four weeks, the overwhelming growth of microorganisms had impoverished the essential nutrients meant for BSFL growth. The dissolved organic compounds were depleted significantly, and because of that, the BSFL had to compete with microorganisms for common growth nutrients. Also, the microorganisms were protected by the strong cell walls or membranes that would forestall valorization by BSFL digestion (Leong et al., 2016). Thus, it is crucial to control the *ex-situ* fermentation

activities to ensure the generated nutritive byproducts from fermentation to spur the BSFL growth will not be depleted by the unnecessary extension of fermentation time.

The *S. cerevisiae* yeast was employed by Li et al. (2015) to execute *ex-situ* fermentation in lignocellulosic biomass of rice straw at 37 °C for 48 hours to improve its protein content prior to BSFL feeding in producing larval biodiesel. The results showed that 89.6% of protein in BSFL feeding substrate could be assimilated into larval biomass within 14 days. The protein-rich fermented rice straw could enrich the accumulation of larval body lipid. Essentially, in the presence of sufficient digestible proteins, the BSFL could synthesize and excrete cellulases enzyme to convert lignocellulosic biomass into its fats and oils. And so, 5.2 g of total lipid could be extracted from the 200 mature BSFL/batch while later producing 4.3 g of biodiesel. This vindicated that the BSFL fed with microbial treated rice straw that consisted mainly of lignocelluloses could be potentially exploited as a feedstock for producing biodiesel while tapping into *ex-situ* fermentation process (Li et al., 2015).

Next, Gao et al. (2019) had assessed the bioconversion performance of BSFL in assimilating *ex-situ* fermented maize straw initially inoculated with *Aspergillus oryzae* fungus at 27°C for 24 hours. The fermented maize straw was more palatable to BSFL since the lignocellulosic content had been hydrolyzed into a more digestible composition as opposed to the untreated maize straw (Binod et al., 2010; Gao et al., 2019; Ware et al., 2005; Zheng et al., 2012a). Nevertheless, in comparing with the commercial wheat bran used as a reference, the feeding with fermented maize straw had led to smaller harvested BSFL, i.e., 1.49 mg/larva against 2.22 mg/larva, while requiring longer rearing duration. The protein content was found to be lower in fermented maize straw than wheat bran, whilst the fermented maize straw was still consisting of higher cellulosic content than the wheat bran, derailing the growth of BSFL (Gao et al., 2019). The BSFL fed with fermented maize straw possessed a lower proportion of saturated fatty acids (45.41%) than the BSFL fed with wheat bran (62.28%). Moreover, the lipid of BSFL fed with fermented maize straw diet had high proportions of monounsaturated fatty acids (24.86%) and polyunsaturated fatty acids (25.37%) which are significant for human health (Calder and Grimble, 2002; Sahena et al., 2009). Moreover, the BSFL fed with fermented maize straw also had a protein content (41.8%) comparable with the conventional soybean meal or aquatic meal (42.1%). Indeed, the biomass from BSFL fed with *ex-situ* fermented maize straw had adequate crude fiber (30.6%) which was usually lacking in other animal feeds (Makkar et al., 2014).

The *ex-situ* fermentation of waste coconut endosperm completed by mixed-bacterial powder (Reckitt Benckiser, UPN:1920080310) for 28 days prior to feeding to BSFL had been

studied by Wong et al. (2019). The waste-to-biomass conversion (WBC) and protein conversion by BSFL were increasing with increasing of mixed-bacterial powder concentrations, reaching the highest values of approximately 9% for WBC and 60% for protein conversion with the initial inoculation concentration of 0.5 wt%. The addition of mixed-bacterial powder exceeding 0.5 wt% during the *ex-situ* fermentation had resulted in the descent of WBC and protein conversion since more microorganisms were competing with BSFL for common nutrients. Moreover, the effect of *ex-situ* fermentation time using the optimum concentration of mixed-bacterial powder, i.e., 0.5 wt%, was also investigated by Wong et al. (2019). In this regard, the best time frame to attain the maximum WBC and protein conversion concurrently was found to be 14 days. By prolonging the time frame beyond 14 days, e.g., 21 and 28 days in a similar study, had brought about no significant impact on BSFL growth and organic waste reduction. From the *ex-situ* fermentation process, the organic acids were found being exuded from the fermented waste coconut endosperm in which were essential for the larval gut health and development (Upadhaya et al., 2016; Wong et al., 2019). Although the addition of mixed-bacterial powder could aid the digestion of fibers from waste coconut endosperm into organic acids, amino acids and vitamins, the significant influence of high amount of celluloses or polymer structures coupled by the low protein content in waste coconut endosperm, i.e., merely 5.83% of protein constituent, had limited the BSFL growth (Caruso et al., 2014; Wong et al., 2019). To epitomize, depending on the type of BSFL feeding substrates, blending method for co-conversion may be somehow more effective than fermentation. Also, the *ex-situ* fermentation approach is deemed time consuming as certain period of time has to be earmarked for the inoculated exo-microbes to complete the fermentation process.

5. Application and limitation of harvested BSFL biomass

In addition to valorizing the solid organic wastes, the harvested BSFL biomass contains valuable biochemical compounds such as lipid, protein, chitin and myriad essential organic minerals that can be potentially employed for alimentation of farmed animals. Also, the larval lipid could be a promising solution serving as the new and sustainable feedstock for biofuel industries, in which various methods to optimize the transesterification process of larval lipid have been currently investigated to maximize and tune the quality of BSFL-based biodiesel (Wong et al., 2019).

The continuous growth of global population and industries has resulted in the rising of fossil fuel consumptions and lately, has been unabatedly dethroned by renewable fuels such

as biodiesel which is non-toxic and eco-friendly toward the environment (Singh et al., 2020). Initially, the biodiesel that is derived from edible crops has led to food shortages. To cushion the menace, biodiesel produced from non-edible crops such as exploiting the spent cooking oil has been proven feasible. However, the operations for converting the spent cooking oil into biodiesel require a high investment cost since large amounts of contaminated matters are present in spent cooking oil, leading to the complication of chemical processes (Mohd-Noor et al., 2017; Tan et al., 2015). Next, the oleaginous microorganisms have been considered as a third generation of biofuel feedstock to produce biodiesel. Alas, the lipid-rich oleaginous microorganisms are experiencing high buoyancy and resisting from settling, incurring a high harvesting cost since extensive time and intensive energy are needed to separate the microalgal biomass from its large cultivation volume (Gerardo et al., 2015; Mohd-Noor et al., 2017; Pinzi et al., 2014). To circumvent these setbacks, the utilization of lipid in the form of fat body from BSFL for the production of biodiesel is gaining more attentions among researchers as a new generation of biofuel feedstock (Cheng and Timilsina, 2011; Manzano-Agugliaro et al., 2012; Mohd-Noor et al., 2017; Payne et al., 2016). The BSFL lipid had been found to possess a higher amount of saturated fats (67%) in comparison with soybean oil (11%) and palm oil (37%) (Hasnol et al., 2020). As the general composition of biodiesel is fatty acid methyl esters (FAMES) mixture, Ushakova et al. (2016) had reported that the FAME profile from BSFL was loaded with C12:0 at 38.43 wt%, followed by C16:1 at 15.71 wt%, C14:0 at 12.33 wt%, C18:1 at 8.81 wt% and C18:0 at 2.95 wt%. Corresponding with Leong et al. (2016) and Wong et al. (2019) studies had also presented that the major composition of FAMES in biodiesel produced from BSFL biomass was lauric acid (C12:0). Surendra et al. (2016) had found the C12:0 in FAMES of biodiesel derived from BSFL reached the peak of 44.9% as opposed to soybean and palm oil-based biodiesel, namely, negligible and 0.1%, respectively. The high level of C12:0 content derived from BSFL lipid indicates the high quality of biodiesel with low viscosity whilst being more stable (Hasnol et al., 2020). Moreover, Zheng et al. (2012a) had confirmed the suitable properties of biodiesel derive from BSFL fed with rice straw and restaurant waste which was in conformity with the requirements of EN 14214 standard. Also, the quality was as well comparable with the biodiesel produced from rapeseed oil (Li et al., 2011).

Apart from being used for biodiesel production, the BSFL lipid is a good fat source for fishmeal in aquaculture. Li et al. (2016c) had proven the lipid from BSFL could substitute the soybean oil as verified in terms of growth of juvenile Jian carp as well as its fatty acid and lipid accumulations during maturity (Wong et al., 2019). The results showed that C12:0 and

C14:0 contents in muscle of experimental fishes were higher when fed with BSFL lipid than soybean oil, *viz.*, 0.49% and 1.30%, respectively, with soybean oil and 4.37% and 2.65%, respectively, with 100% substitution of soybean oil by BSFL lipid in Jian carp diet. However, the growth rates of fishes were not significantly affected by either soybean oil or BSFL lipid, *viz.*, 3.36% and 3.28% of specific growth rates, respectively. This had confirmed the potentiality of BSFL lipid to be adulterated into fishmeal, serving as an alternative composition to soybean oil. In addition, as the high level of medium-chain fatty acids especially C12:0 content in BSFL lipid is similar to that of coconut oil, Kim et al. (2020) proved that the BSFL lipid could significantly increase the unsaturated fatty acids, *i.e.*, linolenic acid, in which resulted in the abundant of omega-3 fatty acids presented in edible chicken meats in comparison with coconut oil employment. Furthermore, the medium-chain fatty acids in BSFL lipid could promote the antibacterial activity and spur the growth performance of broiler chickens when it was laden as a lipid source in the chicken feed (Kim et al., 2020; Li et al., 2016c; Schiavone et al., 2018; Ushakova et al., 2016).

Upon extracting the lipid from BSFL biomass, the residual biomass with concentrated protein source can be subsequently introduced into animal diet (Wong et al., 2019). However, the chitin content in BSFL biomass residues should be degraded by chitinase or removed prior to the utilization since the chitin may retard the growth performances and nutrients adsorption by aquatic farmed fishes (Lindsay, 1984; Makkar et al., 2014; Spranghers et al., 2017; Wong et al., 2019). Li et al. (2017) had studied the replacement of fishmeal protein by defatted BSFL biomass that contained merely 56.9% of crude protein. The results presented that the specific growth rates of Jian carp were not significantly different between using fishmeal and defatted BSFL biomass as the Jian carp feed. Lock et al. (2016) had also proven the potential of replacing fishmeal with BSFL meal and brought about no significant impact on the growth performances of Atlantic salmon. The nutritional compositions of Jian carp fed with either fishmeal or defatted BSFL meal were both fallen within 19-20% for whole body of Jian carp for crude protein, 74-75% for whole body of Jian carp for moisture and about 16% for whole body of Jian carp for lipid, heralding no difference (Li et al., 2017). To top it off, the high catalase activity in dietary defatted BSFL biomass was found contributing into boosting the antioxidant property in Jian carp. Nevertheless, the optimum proportion for substitution of defatted BSFL biomass was recommended to be 50 wt%, since the further increase of defatted BSFL biomass, *i.e.* 75 wt%, could damage the histopathological intestine and contribute to dietary stress (Li et al., 2017). Katya et al. (2017) had also documented that the percentage of BSFL to replace fishmeal or soybean meal as a protein source for juvenile

barramundi should be lesser than 50 wt% for no adverse effect on the whole body proximate and amino acid compositions of barramundi. The BSFL biomass had also been exploited as a protein source instead of soybean meal, i.e., the crude proteins in bird diet were 184.9 and 185.2 g/kg as-fed basis when using soybean meal and replacement of 25 g/kg of soybean meal with BSFL biomass, respectively, for poultry diet (jumbo quails rearing) (Mbhele et al., 2019). Above all, the utilization of BSFL meal for broiler rearing had been reported could decrease the possibility of metabolic skeletal disorders during the development of bird health (Pieterse et al., 2014). The optimum BSFL level to substitute soybean meal was identified at 54 g/kg, since the further inclusion of BSFL in feed could contribute to depress the overall feed intake and body weight increment of jumbo quails.

Besides larval lipid and protein, other biochemicals that can be extracted from BSFL biomass are minerals, vitamins, chitin, etc. Minerals especially calcium can improve the quality of farmed animal growths since calcium is an essential component for muscle mass, enzymatic activity, neuro-signaling, metabolic reaction, synthesis of proteins, maintenance of osmotic and acidic-alkaline equilibria as well as construction of membranes in animal cells (Shumo et al., 2019). The deficiency of calcium will overall result in skeletal, immune and cardiovascular system disorders, bone loss, growth retardation and abnormal posture (Hafeez et al., 2015). Nevertheless, the presence of excess calcium can become a limitation for exploiting BSFL as an animal feed since it will increase the farmed animals' stomach pH. Thereby, impeding the digestion of consumed feeds by farmed animals. Prolonging the retention of remnant feeds in the animal stomach may lead to the diarrhea and risk of bacterial infections especially piglets (Lawlor et al., 2005; Spranghers et al., 2017). The vitamins from BSFL biomass are also considered essential to be presented in farmed animals' meal since it can strengthen the immune system and assist in the digestion process to produce more energies for metabolism and growth (Shumo et al., 2019). Moreover, Borrelli et al. (2017) had reported that BSFL biomass could be employed as a potential prebiotic for laying hens. The chitin content from BSFL could tweak and eventually balance the microbial communities, leading to the reduction of antibiotics utilization in the poultry industries that usually associating to the adverse effects on human health. In addition, Marono et al. (2017) had also presented that the chitin in BSFL meal could reduce the amount of triglycerides and cholesterol, benefiting the health of laying hens. Chitin from BSFL also found enhancing the eggshell thickness and microbiota diversity values in poultries (Kawasaki et al., 2019). Even though the chitin from BSFL has a general positive effect on poultries, the monogastric animals cannot digest the BSFL chitin easily while subsequently, bring about a negative

effect on protein assimilation (Bovera et al., 2016; Longvah et al., 2011; Sánchez-Muros et al., 2014). Also, Makkar et al. (2014) had revealed that the high level of ash in BSFL meal could threaten the growing animal since it would retard the ingestion process of animals especially monogastrics animals and derail growth. Table 4 summarizes the advantages and disadvantages of utilizing BSFL biomass.

6. Conclusions

Black soldier fly larva (BSFL) has a great potential in waste management since it can valorize various organic wastes and transform into its biomass. The difference in organic wastes to serve as the feeding substrates can overall affect the BSFL biomass content especially larval body protein and lipid. Nevertheless, the presence of excessive lignocellulose in the feeding substrates can also hinder the digestion process of BSFL, inhibiting its development. The lack of essential nutrients in the substrates is usually associated to the small larval size harvested. Hence, blending with other substrates and fermentation by microbes have been investigated recently to fortify the nutritional values of BSFL feeding substrates. Upon reaching maturity, the BSFL are harvest for its valuable biochemical content. The protein from BSFL biomass is usually processed into farmed animal feed to replace or substitute fish meal. Moreover, the high content of C12:0 in FAMES mixture has given rise to a good quality of biodiesel produced from BSFL lipid. Besides protein and lipid sources, chitin and calcium from BSFL feedstock also can be used for the livestock alimentation. However, the presence of excess chitin and calcium need to be monitored constantly to preempt retardation of animal growths. To conclude, the employment of BSFL can sustainably valorize various organic wastes, whilst producing green valuable larval biomass to underpin biofuel and livestock industries that eventually benefiting the environment in a dual way. For future development to employ BSFL in valorizing organic wastes, other low-cost and nutrient-rich substrates such as palm kernel expeller could be explored for blending with recalcitrant organic wastes in spurring the palatability of BSFL. The correlation between nutritional constituents of larval feeding substrates and accumulated biochemicals from harvested BSFL biomasses could be as well statistically studied to optimize the performances of BSFL in bioconverting organic wastes into valuable resources. Last but not least, the plausible applications of fine biochemicals derived from BSFL biomass could be investigated in bringing high values to the new industries.

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References

- Aggelides, S., Londra, P., 2000. Effects of compost produced from town wastes and sewage sludge on the physical properties of a loamy and a clay soil. *Bioresour. Technol.* 71, 253-259.
- Aguirre-Villegas, H.A., Larson, R.A., 2017. Evaluating greenhouse gas emissions from dairy manure management practices using survey data and lifecycle tools. *J. Clean. Prod.* 143, 169-179.
- Ahmad, A., Broce, A., Zurek, L., 2006. Evaluation of significance of bacteria in larval development of *Cochliomyia macellaria* (Diptera: Calliphoridae). *J. Med. Entomol.* 43, 1129-1133.
- Alves, A.V., Sanjinez-Argandoña, E.J., Linzmeier, A.M., Cardoso, C.A.L., Macedo, M.L.R., 2016. Food value of mealworm grown on *Acrocomia aculeata* pulp flour. *PLoS One.* 11.
- Amritha, P., Anilkumar, P., 2016. Development of landscaped landfills using organic waste for sustainable urban waste management. *Procedia. Environ. Sci.* 35, 368-376.
- Anjum, M., Khalid, A., Mahmood, T., Aziz, I., 2016. Anaerobic co-digestion of catering waste with partially pretreated lignocellulosic crop residues. *J. Clean. Prod.* 117, 56-63.
- Attiogbe, F.K., Ayim, N.Y.K., Martey, J., 2019. Effectiveness of black soldier fly larvae in composting mercury contaminated organic waste. *Scientific African* 6, e00205.
- Barragan-Fonseca, K.B., Dicke, M., van Loon, J.J., 2017. Nutritional value of the black soldier fly (*Hermetia illucens* L.) and its suitability as animal feed—a review. *J. Insects as Food Feed.* 3, 105-120.

932 Barragan-Fonseca, K.B., Dicke, M., van Loon, J.J., 2018. Influence of larval density and
 933 dietary nutrient concentration on performance, body protein, and fat contents of black
 934 soldier fly larvae (*Hermetia illucens*). *Entomol. Exp. Appl.* 166, 761-770.

935 Barry, T., 2004. Evaluation of the economic, social, and biological feasibility of
 936 bioconverting food wastes with the black soldier fly (*Hermetia illucens*). University of
 937 North Texas.

938 Beniers, J., Graham, R., 2019. Effect of protein and carbohydrate feed concentrations on the
 939 growth and composition of black soldier fly (*Hermetia illucens*) larvae. *J. Insects as*
 940 *Food Feed.* 5, 193-199.

941 Beylot, A., Muller, S., Descat, M., Ménard, Y., Villeneuve, J., 2018. Life cycle assessment of
 942 the French municipal solid waste incineration sector. *Waste Manage.* 80, 144-153.

943 Binod, P., Sindhu, R., Singhania, R.R., Vikram, S., Devi, L., Nagalakshmi, S., Kurien, N.,
 944 Sukumaran, R.K., Pandey, A., 2010. Bioethanol production from rice straw: an
 945 overview. *Bioresour. Technol.* 101, 4767-4774.

946 Bong, C.P.C., Ho, W.S., Hashim, H., Lim, J.S., Ho, C.S., Tan, W.S.P., Lee, C.T., 2017.
 947 Review on the renewable energy and solid waste management policies towards biogas
 948 development in Malaysia. *Renew. Sust. Energ. Rev.* 70, 988-998.

949 Borrelli, L., Coretti, L., Dipineto, L., Bovera, F., Menna, F., Chiariotti, L., Nizza, A., Lembo,
 950 F., Fioretti, A., 2017. Insect-based diet, a promising nutritional source, modulates gut
 951 microbiota composition and SCFAs production in laying hens. *Sci. Rep.* 7, 1-11.

952 Bosch, G., Zhang, S., Oonincx, D.G., Hendriks, W.H., 2014. Protein quality of insects as
 953 potential ingredients for dog and cat foods. *J. Nutr. Sci.* 3, e29.

954 Bovera, F., Loponte, R., Marono, S., Piccolo, G., Parisi, G., Iaconisi, V., Gasco, L., Nizza,
 955 A., 2016. Use of *Tenebrio molitor* larvae meal as protein source in broiler diet: Effect
 956 on growth performance, nutrient digestibility, and carcass and meat traits. *J. Anim.*
 957 *Sci.* 94, 639-647.

958 Cai, M., Hu, R., Zhang, K., Ma, S., Zheng, L., Yu, Z., Zhang, J., 2018. Resistance of black
 959 soldier fly (Diptera: Stratiomyidae) larvae to combined heavy metals and potential
 960 application in municipal sewage sludge treatment. *Environ. Sci. Pollut. Res.* 25, 1559-
 961 1567.

962 Calder, P., Grimble, R., 2002. Polyunsaturated fatty acids, inflammation and immunity. *Eur.*
 963 *J. Clin. Nutr.* 56, S14-S19.

964 Caruso, D., Devic, E., Subamia, I., Talamond, P., Baras, E., 2014. Technical handbook of
 965 domestication and production of Diptera Black Soldier Fly (BSF), *Hermetia illucens*,
 966 *Stratiomyidae*. PT Penerbit IPB Press, Kampus IPB Taman Kencana Bogor.

967 Casartelli, M., Bonelli, M., Bruno, D., Caccia, S., Sgambetterra, G., Cappellozza, S.,
 968 Tettamanti, G., 2019. Structural and functional characterization of *Hermetia illucens*
 969 larval midgut. *Front. Physiol.* 10, 204.

970 Cheng, H., Xu, W., Liu, J., Zhao, Q., He, Y., Chen, G., 2007. Application of composted
 971 sewage sludge (CSS) as a soil amendment for turfgrass growth. *Ecol. Eng.* 29, 96-
 972 104.

973 Cheng, J.J., Timilsina, G.R., 2011. Status and barriers of advanced biofuel technologies: a
 974 review. *Renew. Energ.* 36, 3541-3549.

975 Čičková, H., Newton, G.L., Lacy, R.C., Kozánek, M., 2015. The use of fly larvae for organic
 976 waste treatment. *Waste Manage.* 35, 68-80.

977 Couth, R., Trois, C., 2009. Comparison of waste management activities across Africa with
 978 respect to carbon emissions, Twelfth International Waste Management and Landfill
 979 Symposium, S. Margherita di Pula, Cagliari, Italy, 5-9.

980 Diener, S., Zurbrügg, C., Tockner, K., 2015. Bioaccumulation of heavy metals in the black
 981 soldier fly, *Hermetia illucens* and effects on its life cycle. *J. Insects as Food Feed.* 1,
 982 261-270.

983 Dierenfeld, E.S., King, J., 2008. Digestibility and mineral availability of Phoenix worms,
 984 *Hermetia illucens*, ingested by mountain chicken frogs, *Leptodactylus fallax*. *J.*
 985 *Herpetol. Med. Surg.* 18, 100-105.

986 Dortmans, B., Diener, S., Verstappen, B., Zurbrügg, C., 2017. Black Soldier Fly Biowaste
 987 Processing—A Step-by-Step Guide; Eawag-Swiss Federal Institute of Aquatic
 988 Science and Technology. Department of Sanitation, Water and Solid Waste for
 989 Development (Sandec): Dübendorf, Switzerland.

990 Douglas, A.E., 2015. Multiorganismal insects: diversity and function of resident
 991 microorganisms. *Annu. Rev. Entomol.* 60, 17-34.

992 EEA Greenhouse gas data viewer, 2014. Archive:Greenhouse gas emissions from waste
 993 disposal. Available on line. https://ec.europa.eu/eurostat/statistics-explained/index.php/Archive:Greenhouse_gas_emissions_from_waste_disposal#:~:text=During%20waste%20incineration%2C%20fossil%20carbon,of%20GHG%20emission%20are%20negligible. Accessed on : 19 July 2020.

- Ellis, C., 2018. World Bank: Global waste generation could increase 70% by 2050. Available on line. <https://www.wastedive.com/news/world-bank-global-waste-generation-2050/533031/>. Accessed on : 27 May 2020.
- Engel, P., Moran, N.A., 2013. The gut microbiota of insects—diversity in structure and function. *FEMS Microbiol. Rev.* 37, 699-735.
- Engineering Articles, 2015. Waste Disposal Methods. Environmental Engineering. Available on line. <http://www.engineeringarticles.org/waste-disposal-methods/>. Accessed on : 27 May 2020.
- FAO, 2003. Faostat- Food and Agriculture Organization of the United Nations FAO Statistical Databases. Available on line. <http://faostat.fao.org/>. Accessed on : 27 May 2020.
- FAO, 2015. Food Loss and Food Waste | FAO | Food and Agriculture Organization of the United Nations. Available on line. <http://www.fao.org/food-loss-and-food-waste/en/>. Accessed on : 27 May 2020.
- Ferronato, N., Torretta, V., 2019. Waste mismanagement in developing countries: A review of global issues. *Int. J. Environ. Res. Public Health.* 16, 1060.
- Fitt, G.P., O'Brien, R., 1985. Bacteria associated with four species of *Dacus* (Diptera: Tephritidae) and their role in the nutrition of the larvae. *Oecologia* 67, 447-454.
- Font-Palma, C., 2019. Methods for the treatment of cattle manure—A review. *J. Carbon Res.* 5, 27.
- Förster, M., Klimpel, S., Mehlhorn, H., Sievert, K., Messler, S., Pfeffer, K., 2007. Pilot study on synanthropic flies (eg *Musca*, *Sarcophaga*, *Calliphora*, *Fannia*, *Lucilia*, *Stomoxys*) as vectors of pathogenic microorganisms. *Parasitol. Res.* 101, 243-246.
- Förster, M., Klimpel, S., Sievert, K., 2009. The house fly (*Musca domestica*) as a potential vector of metazoan parasites caught in a pig-pen in Germany. *Vet. Parasitol.* 160, 163-167.
- Gao, Z., Wang, W., Lu, X., Zhu, F., Liu, W., Wang, X., Lei, C., 2019. Bioconversion performance and life table of black soldier fly (*Hermetia illucens*) on fermented maize straw. *J. Clean. Prod.* 230, 974-980.
- Gerardo, M.L., Van Den Hende, S., Vervaeren, H., Coward, T., Skill, S.C., 2015. Harvesting of microalgae within a biorefinery approach: A review of the developments and case studies from pilot-plants. *Algal. Res.* 11, 248-262.
- Gold, M., Cassar, C.M., Zurbrugg, C., Kreuzer, M., Boulos, S., Diener, S., Mathys, A., 2020. Biowaste treatment with black soldier fly larvae: Increasing performance through the

1031 formulation of biowastes based on protein and carbohydrates. Waste Manage. 102,
1032 319-329.

1033 Gómez-Brandón, M., Juárez, M.F.-D., Domínguez, J., Insam, H., 2013. Animal manures:
1034 Recycling and management technologies. Biomass Now-Cultivation and Utilization;
1035 InTech: Rijeka, Croatia, 237-272.

1036 Hafeez, A., Mader, A., Ruhnke, I., Röhe, I., Boroojeni, F.G., Yousaf, M., Männer, K.,
1037 Zentek, J., 2015. Implication of milling methods, thermal treatment, and particle size
1038 of feed in layers on mineral digestibility and retention of minerals in egg contents.
1039 Poult. Sci. 94, 240-248.

1040 Hasnol, S., Kiatkittipong, K., Kiatkittipong, W., Wong, C.Y., Khe, C.S., Lam, M.K., Show,
1041 P.L., Oh, W.D., Chew, T.L., Lim, J.W., 2020. A Review on Insights for Green
1042 Production of Unconventional Protein and Energy Sources Derived from the Larval
1043 Biomass of Black Soldier Fly. Processes 8, 523.

1044 Hogsette, J., Farkas, R., 2000. Secretophagous and hematophagous higher Diptera.
1045 Contributions to a manual of Palearctic Diptera 1, 769-792.

1046 Hong, J., Chen, Y., Wang, M., Ye, L., Qi, C., Yuan, H., Zheng, T., Li, X., 2017.
1047 Intensification of municipal solid waste disposal in China. Renew. Sust. Energ. Rev.
1048 69, 168-176.

1049 Hoornweg, D., Bhada-Tata, P., 2012. What a waste: a global review of solid waste
1050 management. Available on line. [https://openknowledge.worldbank.org/handle/10986](https://openknowledge.worldbank.org/handle/10986/17388)
1051 /17388. Accessed on : 27 May 2020.

1052 Isibika, A., Vinnerås, B., Kibazohi, O., Zurbrügg, C., Lalander, C., 2019. Pre-treatment of
1053 banana peel to improve composting by black soldier fly (*Hermetia illucens* (L.),
1054 Diptera: Stratiomyidae) larvae. Waste Manage. 100, 151-160.

1055 Kadir, M., Velayutham, S., 1999. The management of municipal wastewater sludge in
1056 Malaysia, Symp. on Sludge Management, 18-19.

1057 Kaltenpoth, M., 2009. Actinobacteria as mutualists: general healthcare for insects? Trends in
1058 microbiology 17, 529-535.

1059 Katya, K., Borsra, M., Ganesan, D., Kuppusamy, G., Herriman, M., Salter, A., Ali, S.A.,
1060 2017. Efficacy of insect larval meal to replace fish meal in juvenile barramundi, *Lates*
1061 *calcarifer* reared in freshwater. Int. Aquat. Res. 9, 303-312.

1062 Kawasaki, K., Hashimoto, Y., Hori, A., Kawasaki, T., Hirayasu, H., Iwase, S.-i., Hashizume,
1063 A., Ido, A., Miura, C., Miura, T., 2019. Evaluation of black soldier fly (*Hermetia*

illucens) larvae and pre-pupae raised on household organic waste, as potential ingredients for poultry feed. *Animals* 9, 98.

Kaza, S., Yao, L., Bhada-Tata, P., Van Woerden, F., 2018. What a waste 2.0: a global snapshot of solid waste management to 2050. The World Bank. Available on line. <http://datatopics.worldbank.org/what-a-waste/>. Accessed on : 27 May 2020.

Kim, Y.B., Kim, D.H., Jeong, S.B., Lee, J.W., Kim, T.H., Lee, H.G., Lee, K.W., 2020. Black soldier fly larvae oil as an alternative fat source in broiler nutrition. *Poult. Sci.*, doi : doi.org/10.1016/j.psj.2020.1001.1018.

Kinasih, I., Putra, R.E., Permana, A.D., Gusmara, F.F., Nurhadi, M.Y., Anitasari, R.A., 2018. Growth Performance of Black Soldier Fly Larvae (*Hermetia illucens*) Fed on Some Plant Based Organic Wastes. *Hayati*. 25, 79.

Kirchmann, H., Bernal, M., 1997. Organic waste treatment and C stabilization efficiency. *Soil Biol. Biochem.* 29, 1747-1753.

Kirchmann, H., Lundvall, A., 1993. Relationship between N immobilization and volatile fatty acids in soil after application of pig and cattle slurry. *Biol. Fertil. Soils*. 15, 161-164.

Lalander, C., Diener, S., Zurbrügg, C., Vinnerås, B., 2019. Effects of feedstock on larval development and process efficiency in waste treatment with black soldier fly (*Hermetia illucens*). *J. Clean. Prod.* 208, 211-219.

Laughton, A.M., Garcia, J.R., Altincicek, B., Strand, M.R., Gerardo, N.M., 2011. Characterisation of immune responses in the pea aphid, *Acyrtosiphon pisum*. *J. Insect. Physiol.* 57, 830-839.

Lawlor, P.G., Lynch, P.B., Caffrey, P.J., O'Reilly, J.J., O'Connell, M.K., 2005. Measurements of the acid-binding capacity of ingredients used in pig diets. *Ir. Vet. J.* 58, 447.

Lee, K.P., Simpson, S.J., Raubenheimer, D., 2004. A comparison of nutrient regulation between solitary and gregarious phases of the specialist caterpillar, *Spodoptera exempta* (Walker). *J. Insect. Physiol.* 50, 1171-1180.

Leong, S.Y., Kutty, S.R.M., Malakahmad, A., Tan, C.K., 2016. Feasibility study of biodiesel production using lipids of *Hermetia illucens* larva fed with organic waste. *Waste Manage.* 47, 84-90.

Li, B., Lu, F., Nan, H., Liu, Y., 2012. Isolation and structural characterisation of okara polysaccharides. *Molecules* 17, 753-761.

Li, L., Stasiak, M., Li, L., Xie, B., Fu, Y., Gidzinski, D., Dixon, M., Liu, H., 2016a. Rearing *Tenebrio molitor* in BLSS: Dietary fiber affects larval growth, development, and respiration characteristics. *Acta. Astronaut.* 118, 130-136.

1098 Li, Q., Zheng, L., Qiu, N., Cai, H., Tomberlin, J.K., Yu, Z., 2011. Bioconversion of dairy
1099 manure by black soldier fly (Diptera: Stratiomyidae) for biodiesel and sugar
1100 production. *Waste Manage.* 31, 1316-1320.

1101 Li, R., Chen, S., Li, X., Saifullah Lar, J., He, Y., Zhu, B., 2009. Anaerobic codigestion of
1102 kitchen waste with cattle manure for biogas production. *Energy Fuels.* 23, 2225-2228.

1103 Li, S., Chen, Y., Li, K., Lei, Z., Zhang, Z., 2016b. Characterization of physicochemical
1104 properties of fermented soybean curd residue by *Morchella esculenta*. *Int. Biodeterior.*
1105 *Biodegradation.* 109, 113-118.

1106 Li, S., Ji, H., Zhang, B., Tian, J., Zhou, J., Yu, H., 2016c. Influence of black soldier fly
1107 (*Hermetia illucens*) larvae oil on growth performance, body composition, tissue fatty
1108 acid composition and lipid deposition in juvenile Jian carp (*Cyprinus carpio* var. Jian).
1109 *Aquaculture* 465, 43-52.

1110 Li, S., Ji, H., Zhang, B., Zhou, J., Yu, H., 2017. Defatted black soldier fly (*Hermetia illucens*)
1111 larvae meal in diets for juvenile Jian carp (*Cyprinus carpio* var. Jian): Growth
1112 performance, antioxidant enzyme activities, digestive enzyme activities, intestine and
1113 hepatopancreas histological structure. *Aquaculture* 477, 62-70.

1114 Li, S., Zhu, D., Li, K., Yang, Y., Lei, Z., Zhang, Z., 2013. Soybean curd residue:
1115 composition, utilization, and related limiting factors. *ISRN Industrial Engineering*
1116 2013, doi : [dx.doi.org/10.1155/2013/423590](https://doi.org/10.1155/2013/423590).

1117 Li, W., Li, M., Zheng, L., Liu, Y., Zhang, Y., Yu, Z., Ma, Z., Li, Q., 2015. Simultaneous
1118 utilization of glucose and xylose for lipid accumulation in black soldier fly.
1119 *Biotechnol. Biofuels.* 8, 117.

1120 Lim, J.W., Mohd-Noor, S.N., Wong, C.Y., Lam, M.K., Goh, P.S., Beniers, J., Oh, W.D.,
1121 Jumbri, K., Ghani, N.A., 2019. Palatability of black soldier fly larvae in valorizing
1122 mixed waste coconut endosperm and soybean curd residue into larval lipid and
1123 protein sources. *J. Environ. Manage.* 231, 129-136.

1124 Lim, S.L., Lee, L.H., Wu, T.Y., 2016. Sustainability of using composting and
1125 vermicomposting technologies for organic solid waste biotransformation: recent
1126 overview, greenhouse gases emissions and economic analysis. *J. Clean. Prod.* 111,
1127 262-278.

1128 Lindsay, G., 1984. Distribution and function of digestive tract chitinolytic enzymes in fish. *J.*
1129 *Fish. Biol.* 24, 529-536.

1130 Liyala, C.M., 2011. Modernising solid waste management at municipal level: Institutional
1131 arrangements in urban centres of East Africa. *Wageningen Academic Pub.* 3, 174.

1132 Lock, E., Arsiwalla, T., Waagbø, R., 2016. Insect larvae meal as an alternative source of
 1133 nutrients in the diet of Atlantic salmon (*Salmo salar*) postsmolt. *Aquac. Nutr.* 22,
 1134 1202-1213.

1135 Longvah, T., Mangthya, K., Ramulu, P., 2011. Nutrient composition and protein quality
 1136 evaluation of eri silkworm (*Samia ricinii*) prepupae and pupae. *Food Chem.* 128, 400-
 1137 403.

1138 Loyon, L., 2018. Overview of Animal Manure Management for Beef, Pig, and Poultry Farms
 1139 in France. *Front. Sustain. Food. Syst.* 2, 36.

1140 Makkar, H.P., Tran, G., Heuzé, V., Ankers, P., 2014. State-of-the-art on use of insects as
 1141 animal feed. *Anim. Feed. Sci. Technol.* 197, 1-33.

1142 Malek, M.A., Shaaban, M., 2008. Landfill Common Method and Practices of Solid Waste
 1143 Disposal in Malaysia. *ISWA World Congr.* MYS100320081047.

1144 Malomo, G.A., Madugu, A.S., Bolu, S.A., 2018. Sustainable Animal Manure Management
 1145 Strategies and Practices. *Agricultural Waste and Residues*, 119. doi :
 1146 110.5772/intechopen.78645.

1147 Manzano-Agugliaro, F., Sanchez-Muros, M., Barroso, F., Martínez-Sánchez, A., Rojo, S.,
 1148 Pérez-Bañón, C., 2012. Insects for biodiesel production. *Renew. Sust. Energ. Rev.* 16,
 1149 3744-3753.

1150 Marono, S., Loponte, R., Lombardi, P., Vassalotti, G., Pero, M., Russo, F., Gasco, L., Parisi,
 1151 G., Piccolo, G., Nizza, S., 2017. Productive performance and blood profiles of laying
 1152 hens fed *Hermetia illucens* larvae meal as total replacement of soybean meal from 24
 1153 to 45 weeks of age. *Poult. Sci.* 96, 1783-1790.

1154 Masih-Das, J., Tao, W., 2018. Anaerobic co-digestion of foodwaste with liquid dairy manure
 1155 or manure digestate: Co-substrate limitation and inhibition. *J. Environ. Manage.* 223,
 1156 917-924.

1157 Mata-Alvarez, J., Dosta, J., Romero-Güiza, M., Fonoll, X., Peces, M., Astals, S., 2014. A
 1158 critical review on anaerobic co-digestion achievements between 2010 and 2013.
 1159 *Renew. Sust. Energ. Rev.* 36, 412-427.

1160 Mateo-Sagasta, J., Raschid-Sally, L., Thebo, A., 2015. Global wastewater and sludge
 1161 production, treatment and use, *Wastewater*. Springer, 15-38.

1162 Mbhele, F.G., Mnisi, C.M., Mlambo, V., 2019. A Nutritional Evaluation of Insect Meal as a
 1163 Sustainable Protein Source for Jumbo Quails: Physiological and Meat Quality
 1164 Responses. *Sustainability* 11, 6592.

1165 Mohd-Noor, S.N., Wong, C.Y., Lim, J.W., Uemura, Y., Lam, M.K., Ramli, A., Bashir,
 1166 M.J., Tham, L., 2017. Optimization of self-fermented period of waste coconut
 1167 endosperm destined to feed black soldier fly larvae in enhancing the lipid and protein
 1168 yields. *Renew. Energ.* 111, 646-654.

1169 Move for Hunger, 2015. The Environmental Impact of Food Waste. Food Waste. Available
 1170 on line. <https://moveforhunger.org/the-environmental-impact-of-food-waste>.
 1171 Accessed on : 1 April 2020.

1172 National Bureau of Statistics of the People's Republic of China, 2015. China Statistic
 1173 Yearbook 2015. Available on line. [http://www.stats.gov.cn/tjsj/ndsj/2015/](http://www.stats.gov.cn/tjsj/ndsj/2015/indexch.htm)
 1174 [indexch.htm](http://www.stats.gov.cn/tjsj/ndsj/2015/indexch.htm). Accessed on : 1 May 2020.

1175 Newton, G., Sheppard, D., Watson, D., Burtle, G., Dove, C., Tomberlin, J., Thelen, E., 2005.
 1176 The black soldier fly, *Hermetia illucens*, as a manure management/resource recovery
 1177 tool, Symposium on the state of the science of Animal Manure and Waste
 1178 Management. *Semantic Scholar*, 5-7.

1179 Nguyen, T.T., Tomberlin, J.K., Vanlaerhoven, S., 2013. Influence of resources on *Hermetia*
 1180 *illucens* (Diptera: Stratiomyidae) larval development. *J. Med. Entomol.* 50, 898-906.

1181 Nijhout, H., 2003. The control of body size in insects. *Developmental biology* 261, 1-9.

1182 Norgren, R., Björkqvist, O., Jonsson, A., 2019. Bio-sludge from the Pulp and Paper Industry
 1183 as Feed for Black Soldier Fly Larvae: A Study of Critical Factors for Growth and
 1184 Survival. *Waste. Biomass. Valorization.*, 1-7.

1185 Oladejo, J., Shi, K., Luo, X., Yang, G., Wu, T., 2019. A review of sludge-to-energy recovery
 1186 methods. *Energies.* 12, 60.

1187 Oonincx, D., Van Huis, A., Van Loon, J., 2015. Nutrient utilisation by black soldier flies fed
 1188 with chicken, pig, or cow manure. *J. Insects as Food Feed.* 1, 131-139.

1189 Pace, S.A., Yazdani, R., Kendall, A., Simmons, C.W., VanderGheynst, J.S., 2018. Impact of
 1190 organic waste composition on life cycle energy production, global warming and
 1191 Water use for treatment by anaerobic digestion followed by composting. *Resour.*
 1192 *Conserv. Recycl.* 137, 126-135.

1193 Payne, C.L., Dobermann, D., Forkes, A., House, J., Josephs, J., McBride, A., Müller, A.,
 1194 Quilliam, R., Soares, S., 2016. Insects as food and feed: European perspectives on
 1195 recent research and future priorities. *J. Insects as Food Feed.* 2, 269-276.

1196 Pieterse, E., Pretorius, Q., Hoffman, L., Drew, D., 2014. The carcass quality, meat quality
 1197 and sensory characteristics of broilers raised on diets containing either *Musca*

domestica larvae meal, fish meal or soya bean meal as the main protein source. Anim. Prod. Sci. 54, 622-628.

Pinzi, S., Leiva, D., López-García, I., Redel-Macías, M.D., Dorado, M.P., 2014. Latest trends in feedstocks for biodiesel production. Biofuel. Bioprod. Biorefin. 8, 126-143.

Popa, R., Green, T.R., 2012. Using black soldier fly larvae for processing organic leachates. J. Econ. Entomol. 105, 374-378.

Ramos-Bueno, R.P., González-Fernández, M.J., Sánchez-Muros-Lozano, M.J., García-Barroso, F., Guil-Guerrero, J.L., 2016. Fatty acid profiles and cholesterol content of seven insect species assessed by several extraction systems. Eur. Food Res. Technol. 242, 1471-1477.

Recycle Bank, 2006. The impact of organic waste. Available on line. <https://visual.ly/community/infographic/animals/impact-organic-waste>. Accessed on : 27 May 2020.

Roslan, S.N., Ghazali, S.S., Asli, N.M., 2013. Study on the characteristics and utilization of sewage sludge at Indah Water Konsortium (IWK) Sungai Udang, Melaka, Proceedings of World Academy of Science, Engineering and Technology. World Academy of Science, Engineering and Technology (WASET), 647.

Sahena, F., Zaidul, I., Jinap, S., Saari, N., Jahurul, H., Abbas, K., Norulaini, N., 2009. PUFAs in fish: extraction, fractionation, importance in health. Compr. Rev. Food. Sci. Food. Saf. 8, 59-74.

Sánchez-Muros, M.-J., Barroso, F.G., Manzano-Agugliaro, F., 2014. Insect meal as renewable source of food for animal feeding: a review. J. Clean. Prod. 65, 16-27.

Sauve, G., Van Acker, K., 2020. The environmental impacts of municipal solid waste landfills in Europe: A life cycle assessment of proper reference cases to support decision making. J. Environ. Manage. 261, 110216.

Schiavone, A., Dabbou, S., De Marco, M., Cullere, M., Biasato, I., Biasibetti, E., Capucchio, M., Bergagna, S., Dezzutto, D., Meneguz, M., 2018. Black soldier fly larva fat inclusion in finisher broiler chicken diet as an alternative fat source. Animal 12, 2032-2039.

Scott, J.J., Oh, D.C., Yuceer, M.C., Klepzig, K.D., Clardy, J., Currie, C.R., 2008. Bacterial protection of beetle-fungus mutualism. Science 322, 63-63.

Sheppard, D.C., Newton, G.L., Thompson, S.A., Savage, S., 1994. A value added manure management system using the black soldier fly. Bioresour. Technol. 50, 275-279.

1231 Shumo, M., Osuga, I.M., Khamis, F.M., Tanga, C.M., Fiaboe, K.K., Subramanian, S., Ekesi,
1232 S., van Huis, A., Borgemeister, C., 2019. The nutritive value of black soldier fly
1233 larvae reared on common organic waste streams in Kenya. *Sci. Rep.* 9, 1-13.

1234 Simpson, S.J., Sword, G.A., Lorch, P.D., Couzin, I.D., 2006. Cannibal crickets on a forced
1235 march for protein and salt. *Proceedings of the National Academy of Sciences* 103,
1236 4152-4156.

1237 Singh, D., Sharma, D., Soni, S., Sharma, S., Sharma, P.K., Jhalani, A., 2020. A review on
1238 feedstocks, production processes, and yield for different generations of biodiesel. *Fuel*
1239 262, 116553.

1240 Somroo, A.A., ur Rehman, K., Zheng, L., Cai, M., Xiao, X., Hu, S., Mathys, A., Gold, M.,
1241 Yu, Z., Zhang, J., 2019. Influence of *Lactobacillus buchneri* on soybean curd residue
1242 co-conversion by black soldier fly larvae (*Hermetia illucens*) for food and feedstock
1243 production. *Waste Manage.* 86, 114-122.

1244 Sprangers, T., Ottoboni, M., Klootwijk, C., Oryn, A., Deboosere, S., De Meulenaer, B.,
1245 Michiels, J., Eeckhout, M., De Clercq, P., De Smet, S., 2017. Nutritional composition
1246 of black soldier fly (*Hermetia illucens*) prepupae reared on different organic waste
1247 substrates. *J. Sci. Food Agric.* 97, 2594-2600.

1248 Steinfeld, H., Gerber, P., Wassenaar, T., Castel, V., Rosales, M., Rosales, M., de Haan, C.,
1249 2006. *Livestock's long shadow: environmental issues and options.* Food &
1250 Agriculture Org, pp. 1-390.

1251 Surendra, K., Olivier, R., Tomberlin, J.K., Jha, R., Khanal, S.K., 2016. Bioconversion of
1252 organic wastes into biodiesel and animal feed via insect farming. *Renew. Energ.* 98,
1253 197-202.

1254 Szogi, A.A., Vanotti, M.B., Ro, K.S., 2015. Methods for treatment of animal manures to
1255 reduce nutrient pollution prior to soil application. *Curr. Pollut. Rep.* 1, 47-56.

1256 Tan, C.H., Show, P.L., Chang, J.-S., Ling, T.C., Lan, J.C.-W., 2015. Novel approaches of
1257 producing bioenergies from microalgae: A recent review. *Biotechnol. Adv.* 33, 1219-
1258 1227.

1259 Teixeira, L., Ferreira, Á., Ashburner, M., 2008. The bacterial symbiont *Wolbachia* induces
1260 resistance to RNA viral infections in *Drosophila melanogaster*. *PLoS Biol.* 6.

1261 Tomberlin, J., Cammack, J., 2017. Black soldier fly: biology and mass production. *Insects as*
1262 *food and feed: from production to consumption.* Wageningen Academic Publishers,
1263 Wageningen, the Netherlands, 231-246.

1264 Tschirner, M., Simon, A., 2015. Influence of different growing substrates and processing on
1265 the nutrient composition of black soldier fly larvae destined for animal feed. *J. Insects*
1266 as Food Feed. 1, 249-259.

1267 Upadhaya, S.D., Lee, K.Y., Kim, I.H., 2016. Effect of protected organic acid blends on
1268 growth performance, nutrient digestibility and faecal micro flora in growing pigs. *J.*
1269 *Appl. Anim. Res.* 44, 238-242.

1270 ur Rehman, K., Cai, M., Xiao, X., Zheng, L., Wang, H., Soomro, A.A., Zhou, Y., Li, W., Yu,
1271 Z., Zhang, J., 2017a. Cellulose decomposition and larval biomass production from the
1272 co-digestion of dairy manure and chicken manure by mini-livestock (*Hermetia*
1273 *illucens* L.). *J. Environ. Manage.* 196, 458-465.

1274 ur Rehman, K., Rehman, A., Cai, M., Zheng, L., Xiao, X., Somroo, A.A., Wang, H., Li, W.,
1275 Yu, Z., Zhang, J., 2017b. Conversion of mixtures of dairy manure and soybean curd
1276 residue by black soldier fly larvae (*Hermetia illucens* L.). *J. Clean. Prod.* 154, 366-
1277 373.

1278 ur Rehman, K., Rehman, R.U., Somroo, A.A., Cai, M., Zheng, L., Xiao, X., Rehman, A.U.,
1279 Rehman, A., Tomberlin, J.K., Yu, Z., 2019. Enhanced bioconversion of dairy and
1280 chicken manure by the interaction of exogenous bacteria and black soldier fly larvae.
1281 *J. Environ. Manage.* 237, 75-83.

1282 Ushakova, N., Brodskii, E., Kovalenko, A., Bastrakov, A., Kozlova, A., Pavlov, D., 2016.
1283 Characteristics of lipid fractions of larvae of the black soldier fly *Hermetia illucens*,
1284 *Doklady biochemistry and biophysics*. Springer, 209-212.

1285 Veldkamp, T., Van Duinkerken, G., van Huis, A., Lakemond, C., Ottevanger, E., Bosch, G.,
1286 Van Boekel, T., 2012. Insects as a Sustainable Feed Ingredient in Pig and Poultry
1287 Diets: a Feasibility Study= Insecten als duurzame diervoedergrondstof in varkens-en
1288 pluimveevoeders: een haalbaarheidsstudie. Wageningen UR Livestock Research, 1-
1289 345.

1290 Wang, D., Ai, J., Shen, F., Yang, G., Zhang, Y., Deng, S., Zhang, J., Zeng, Y., Song, C.,
1291 2017a. Improving anaerobic digestion of easy-acidification substrates by promoting
1292 buffering capacity using biochar derived from vermicompost. *Bioresour. Technol.*
1293 227, 286-296.

1294 Wang, H., ur Rehman, K., Liu, X., Yang, Q., Zheng, L., Li, W., Cai, M., Li, Q., Zhang, J.,
1295 Yu, Z., 2017b. Insect biorefinery: a green approach for conversion of crop residues
1296 into biodiesel and protein. *Biotechnol. Biofuels.* 10, 304.

1297 Wang, Y., Zhang, X., Liao, W., Wu, J., Yang, X., Shui, W., Deng, S., Zhang, Y., Lin, L.,
1298 Xiao, Y., 2018. Investigating impact of waste reuse on the sustainability of municipal
1299 solid waste (MSW) incineration industry using emergy approach: A case study from
1300 Sichuan province, China. *Waste Manage.* 77, 252-267.

1301 Ware, R., Torrentera, N., Zinn, R., 2005. Influence of maceration and fibrolytic enzymes on
1302 the feeding value of rice straw. *J. Anim. Vet. Adv.*, 387-392.

1303 Wiedmeier, R., Arambel, M., Walters, J., 1987. Effect of yeast culture and *Aspergillus oryzae*
1304 fermentation extract on ruminal characteristics and nutrient digestibility. *J. Dairy. Sci.*
1305 70, 2063-2068.

1306 Wilson, L., 2012. The food wastage footprint is big. Globalisation and food consumption,
1307 Today's world. Available on line. [http://shrinkthatfootprint.com/the-big-footprint-of-](http://shrinkthatfootprint.com/the-big-footprint-of-food-waste)
1308 food-waste. Accessed on : 1 April 2020.

1309 Wong, C.Y., Rosli, S.S., Uemura, Y., Ho, Y.C., Leejeerajumnean, A., Kiatkittipong, W.,
1310 Cheng, C.-K., Lam, M.-K., Lim, J.-W., 2019. Potential Protein and Biodiesel Sources
1311 from Black Soldier Fly Larvae: Insights of Larval Harvesting Instar and Fermented
1312 Feeding Medium. *Energies.* 12, 1570.

1313 Wong, C.Y., Mohd Aris, M.N., Daud, H., Lam, M.K., Yong, C.S., Abu Hasan, H., Chong, S.,
1314 Show, P.L., Hajoeningtijas, O.D., Ho, Y.C., 2020. In-Situ Yeast Fermentation to
1315 Enhance Bioconversion of Coconut Endosperm Waste into Larval Biomass of
1316 *Hermetia illucens*: Statistical Augmentation of Larval Lipid Content. *Sustainability*
1317 12, 1558.

1318 Wright, G.A., Simpson, S.J., Raubenheimer, D., Stevenson, P.C., 2003. The feeding behavior
1319 of the weevil, *Exophthalmus jekelianus*, with respect to the nutrients and
1320 allelochemicals in host plant leaves. *Oikos* 100, 172-184.

1321 Xiao, X., Mazza, L., Yu, Y., Cai, M., Zheng, L., Tomberlin, J.K., Yu, J., van Huis, A., Yu,
1322 Z., Fasulo, S., 2018. Efficient co-conversion process of chicken manure into protein
1323 feed and organic fertilizer by *Hermetia illucens* L.(Diptera: Stratiomyidae) larvae and
1324 functional bacteria. *J. Environ. Manage.* 217, 668-676.

1325 Yoon, S.-H., Mukerjea, R., Robyt, J.F., 2003. Specificity of yeast (*Saccharomyces cerevisiae*)
1326 in removing carbohydrates by fermentation. *Carbohydr. Res.* 338, 1127-1132.

1327 Yu, G., Cheng, P., Chen, Y., Li, Y., Yang, Z., Chen, Y., Tomberlin, J.K., 2011. Inoculating
1328 poultry manure with companion bacteria influences growth and development of black
1329 soldier fly (Diptera: Stratiomyidae) larvae. *Environ. Entomol.* 40, 30-35.

- Yu, G., Nil, C., He, G., Zhou, L., Xia, Q., Cheng, P., 2010. Isolation and identification of bacteria producing enzymes from gut and skin of black soldier fly (*Hermetia illulens*) larvae. *Kun. Chong. Zhi. Shi.* 47, 889-894.
- Yun, J.H., Roh, S.W., Whon, T.W., Jung, M.J., Kim, M.S., Park, D.S., Yoon, C., Nam, Y.D., Kim, Y.J., Choi, J.H., 2014. Insect gut bacterial diversity determined by environmental habitat, diet, developmental stage, and phylogeny of host. *Appl. Environ. Microbiol.* 80, 5254-5264.
- Zhang, H., Schroder, J., 2014. Animal manure production and utilization in the US, *Applied manure and nutrient chemistry for sustainable agriculture and environment.* Springer, 1-21.
- Zhao, H., Themelis, N., Bourtsalas, A., McGillis, W., 2019. Methane Emissions from Landfills. Columbia University.
- Zheng, L., Hou, Y., Li, W., Yang, S., Li, Q., Yu, Z., 2012a. Biodiesel production from rice straw and restaurant waste employing black soldier fly assisted by microbes. *Energy* 47, 225-229.
- Zheng, L., Hou, Y., Li, W., Yang, S., Li, Q., Yu, Z., 2013. Exploring the potential of grease from yellow mealworm beetle (*Tenebrio molitor*) as a novel biodiesel feedstock. *Appl. Energy.* 101, 618-621.
- Zheng, L., Li, Q., Zhang, J., Yu, Z., 2012b. Double the biodiesel yield: Rearing black soldier fly larvae, *Hermetia illucens*, on solid residual fraction of restaurant waste after grease extraction for biodiesel production. *Renew. Energ.* 41, 75-79.